

**STUDY OF CELL MEDIATED  
IMMUNITY IN LEPROSY**

**THESIS  
FOR DOCTOR OF MEDICINE  
[ PATHOLOGY ]**

**BUNDELKHAND UNIVERSITY,  
JHANSI.**



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CERTIFICATE

This is to certify that the work in connection with Thesis "STUDY OF CELL-MEDIATED IMMUNITY IN LEPROSY" for M.B. (Pathology) of Bundelkhand University was conducted in the Department of Pathology by Dr. PREM KUMAR SINGH under my guidance and supervision. The techniques embodied in the thesis were undertaken by the candidate himself and observations recorded have been periodically checked by me.

He has put in the necessary stay in the department according to University regulations.

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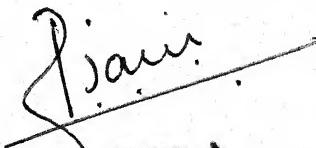
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## ACKNOWLEDGEMENT

\*\*\*\*\*

It is please reverence that I express my profound gratitude to my esteemed teacher Dr. V.P. Mitra, M.B.(Path.), Professor and Head of the Department of Pathology, M.L.B. Medical College, Jhansi, under whose expert and untiring guidance, I had an opportunity to work even at his personal inconvenience. He has been a constant source of encouragement and paternal guidance in my moments of despair. The present study carries at every stage the glittering imprint of his dynamic personality, wise and concrete suggestions, meticulous attention, and mature and reasoned criticism.

I shall ever remain indebted to Dr. Ratna, M.D. (Path.), Lecturer in Department of Pathology, M.L.B. Medical College, Jhansi, for her perfect guidance and untiring help. Her timely suggestions and critical approach during entire course of study, went a long way in materializing this work in its presentable form. Her affectionate nature and indefatigable spirit were constant beacon of moral support to look up during lean moment.

I am equally grateful to Dr. P.K. Jain, M.B., M.M.A.M.B., Lecturer in Medicine, M.L.B. Medical College, Jhansi, for kind and every conceivable help

to me in carrying out the rather extensive clinical work in Department of Skin and V.D.

Words fail to express my feeling of deep respect to Dr. R.K. Gupta, M.D.(Path.), Reader in Department of Pathology, M.L.B. Medical College, Jhansi, for having freely backed upon his profound knowledge and endless resources. Without his co-operation, present study may well not have materialized at all.

I am thankful to Dr. V.K. Sharma, M.D.(Path.), Lecturer in Department of Pathology, M.L.B. Medical College, Jhansi, for his time to time and much solicited help.

I wish to accept my gratitude to Dr. U. Ganguly, Ph.D., Senior Research Officer and Dr. S.K. Chet, Research Officers and others, Central JAIIBA Institute for Leprosy (NHR), Agra, who initiated me in the technical expertise necessary for present study and providing the vital antigen.

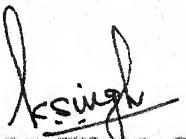
I extend my thanks to Dr. B.L. Verma, Ph.D., Statistician-cum-Lecturer in Department of S.P.M., M.L.B. Medical College, Jhansi, for help in statistical analysis.

Although friends do not need such words, yet I extend my loving and affectionate thanks to all my friends for their co-operation and help whenever required.

My special thanks to Mr. N.S. Saman, for bringing out all the work in the presentable form by his excellent ability of preparing the type script.

I shall also be thankful and full appreciate-  
tion for inconveniences suffered by every patient,  
his/her relatives, and paramedical staff members  
especially Mrs. S. Chahar during the course of  
present study.

Humble shall be I in this moment of glory  
and reward for all the credit accrued to me, shall  
actually go to my parents and family members: for  
without their affection, understanding, suffering,  
and sacrifice, this study may well not have seen the  
light of day at all.



( PREM KUMAR SINGH )

**"Improper work is not merely medical relief, it is transforming the frustration in life into the joy of dedication, personal ambition in to selfless service. If you can transform the life of a patient or change his values of life you can change the village and country".**

**"MAHATMA GANDHI"**

**C O N T E N T S**

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## INTRODUCTION



Leprosy is one of the most challenging disease known today, ranking cancer in its damage and lack of adequate knowledge but even more challenging because of what leprosy does to its victims physically, socio-economically and psychologically.

It is a disease of great antiquity. The leper has for centuries been a social out cast, partly, from biblical times he was regarded as unclean and partly because his repulsive appearance and disabilities prevented him from being an acceptable member of the community.

Hatred and ostracism cause concealment of the disease on the part of patients until it becomes too obvious. In doing so, the sufferer unintentionally helps to exaggerate the disease in himself as he remains without any treatment. Consequently disease takes longer time in becoming acute enough to manifest itself too obviously in the patient. Even by this time he would have infected many other persons in his community.

The total number of leprosy cases in the world are estimated to be approximately 12 million. This figure for India being about 8.2 million patients and it is widely distributed in all parts

of this country. Causative organism of this disease is *Mycobacterium leprae* bacilli which was recognized by Hansen in 1874.

Clinically leprosy is manifested in two main clinical forms, lepromatous leprosy and tuberculoid leprosy and these two types represent the opposite poles - lack of resistance and resistance in host - respectively. Thus realizing the importance of host immune status, an immunological approach may help in proper pathogenesis, diagnosis, control and prevention of leprosy. Ridley and Gough (1966) have provided nomenclature, in the form of a system of diagnostic classification that is fundamental to most current immunological investigations.

As an elaboration of polar concept, Ridley and Jopling (1966) first proposed a system of five numbered classification. They retained the traditional tuberculoid pole (TT), lepromatous pole (LL) and borderline (BB) group, but added two intermediary categories, borderline with tuberculoid features (BT) and borderline with lepromatous features (BL) - TT, BT, BB, BL, LL, comprise a spectrum in continuity. They also explained that each stage in spectrum was determined by the result of host response to antigen of *Mycobacterium leprae*. Patients with LL have more immunity against *Mycobacterium leprae* than do the LL but less

than patients with II and TT. So it indicates that TT patients have highest and LL have lowest immunity.

Landsteiner and Chase (1943) demonstrated that delayed type of hypersensitivity could be conferred on non-reactive subjects by transferring living lymphoid cells from sensitized donors. These observations provided the foundation for science of cellular immunology. The past few years have shown that T cell count, PHA response, response to chemicals such as DNCB and graft transplantation, may help in assessing the cellular immunity.

Present study has been undertaken to assess the cellular immunity in different types of leprosy patients. The tests, which were taken in assessment, were status of T-cell and B-cell in peripheral blood, lymphocyte response to PHA and skin recall test using lepromin and candida antigens.

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## **REVIEW OF LITERATURE**



### HISTORY OF LEPROSY :

Leprosy is a disease of great antiquity; its origin and early spread is, however, largely a matter of surmise. Possibly it originated in Africa and had spread very early to India. Scott (1943) remarked that it was not possible to declare with certainty, in what country leprosy originated but study of available records point out to its first home being Africa.

Dhamondra had quoted that "In Shushrat Samita (600 BC)" one finds a reasonably good account of clinical features and treatment of disease and references of leprosy are made as "Vat-Rakata" or "Vat-Shoulta" and "Kushta" in this book.

If loss of Manus, regarded to contain certain instructions about the prophylaxis of leprosy according to Roger and Muir (1960) and Loss (1942), the disease mentioned in Vedas as "Kushta" will date back to 1400 BC.

Sir William MacArthur pointed out that word leper is derived from a word meaning, a scale or parchment and that the latin word for book-lister has the same derivation (Ogilvie et al, 1936).

Leprosy is mentioned at several places in Bible but it is doubtful whether the words have some reference to the disease leprosy what we know today.

Saroth in Jewish literature and lepro in Arabic literature stand for scaly and fungal diseases. The term Saroth in Old Testament and the term lepro in New Testament have been considered by many authors to refer to leprosy and in all Biblical translations have been rendered as leprosy.

This view however is being challenged by many recent writers including Lie (1930) and Lestrem (1952). Anderson (1969) has reviewed the whole matter carefully and has concluded that there is no evidence of leprosy in Biblical writings. This is probably an unfortunate association that Biblical leprosy and the disease, as is now known, have assumed the same name.

#### PATHOLOGY OF LEPROSY :

It is a fact that leprosy is an infectious disease, caused by *Mycobacterium leprae*. It has attracted and intrigued Immunologists over the last decade for various reasons. Firstly, a large population is exposed to *Myc. leprae* infection and goes through a stage of subclinical infection. Most of these people are able to mount a resistance to *Myc. leprae* infection. A minor group of population, due to unknown mechanism, are unable to mount a proper immune response against *Myc. leprae* invasion. These people manifest variety of clinical symptoms ranging from single, often self healing lesions or tuberculoid leprosy, to a disseminated and progressive disease manifested as lepromatous leprosy.

Secondly, inspite of long generation time of *Mycobacterium leprae*, leprosy may be punctuated by acute exacerbation episodes with features in common with hypersensitivity reactions.

Thirdly there is lack of methods for identification of high risk group and immunoprophylaxis in the leprosy cases (Cedal 1978).

In the great majority, an effective immune response is to arrest the multiplication of *Mycobacterium leprae* at subclinical level and this prevents the development of clinical manifestations. Among those who develop clinical manifestation, immune responses to *Mycobacterium leprae* seem to play a major part in expression of clinical symptoms.

#### I- CHARACTERISATION OF MYCOBACTERIUM LEPRAE ANTIGEN AND SUSPECT MECHANISM IN HOST RESISTANCE TO INTRACELLULAR BACTERIA :

*Mycobacterium leprae* is an obligatory intracellular organism with especial affinity for skin, nerve and muscle tissues. It has very long generation time, somewhere between 10-30 days in comparison to tubercle bacillus which divides about every 20 hours and coliform bacteria every 20 minutes. The leprosy bacillus however, still defies cultivation outside the human body, although the situation has been mitigated to some extent by discovery of amodilite in which

towards anode. The latter component only induces delayed type of hypersensitivity skin reaction in tuberculous patients. This study indicates that specificity of antigen may lie in the latter fraction.

### **III- HISTOPATHOLOGICAL AND CLINICAL CLASSIFICATION**

#### **BASED ON IMMUNITY :**

The spectrum of clinical manifestation caused by infection with *Mycobacterium leprae* include two polar type of infection lepromatous and tuberculoid leprosy; a very broad range of intermediate forms classified as borderline (Arnold 1974, Raballe and Azulay 1978). Ridley and Jopling (1968, 1966) have provided a nomenclature, in the form of system of diagnostic classification, that is fundamental to most current immunological investigations. The following classification was developed as a result of careful correlation of the clinical features of the disease and histopathological pattern in skin biopsies.

**TT      Tuberculoid polar (High resistance form)**

**BT      Borderline tuberculoid.**

**BB      Border line.**

**BL      Borderline lepromatous.**

**LL      Lepromatous polar (low resistance form).**

In study of patients in Malaya, Ridley and Waters (1969) separated a sixth group from the Ridley-Jopling classification that lay between true polar lepromatous and borderline lepromatous; this subpopulation category was named as leprosy indefinite (LI). Patients with LI were found to clear AFB more rapidly than patients with LL.

An analogue tuberculin indefinite (TI) group that lay between TT and BT was defined by Ridley (1972), which completed a seven numbered spectrum - TT, TI, BT, BB, BL, LI and LL in continuity. Myrvang et al (1973) studies Myco-leprose induced lymphocyte transformation and migration-inhibition factor production and found that lymphocyte from patients with LI did not respond in a greatly different manner than from those patients with LL, which suggests that distinction between LL and LI may be spurious immunologically. Rao et al (1976) have shown that cases histologically classifiable as LI have clinical features of LL. Thus histological distinction between LL and LI is without importance.

It is clear from above that five numbered classification of Ridley and Jopling (1966) is useful in clinical application, where clinician can have an idea regarding the extent of disease process

especially when a patient is shifting immunologically either upward or downward in the spectrum. The stage of immunity in patients have been worked out in detail along with Ridley-Jopling scale. It is obvious from studies that as disease glides down gradually in spectrum from tuberculoid to lepromatous end of the pole, there is gradual decrease in cell mediated immunity.

**Variant forms of leprosy not included in Ridley-Jopling scale :**

There are two other important variants of leprosy which are not included in Ridley-Jopling classification.

#### (1) Indeterminate leprosy :

The clinical and histological features are not so distinct in this early stage of disease as to provide definite nature of immunological status. There are one or more hypopigmented macule. The indeterminate lesions may apparently regress spontaneously or progress to become tuberculoid, borderline or lepromatous leprosy or remain as indeterminate over a prolonged period of time. The mechanism behind the hypopigmentation remains unknown. Godal (1975) has quoted the possibility that Myco. leprae itself interferes with pigment

production. Another is that nonspecific infiltration of inflammatory cells suppresses pigment production. The fact that *Mycobacterium leprae* may be found in large quantities in lepromatous patients without hypopigmentation favours the last hypothesis.

### (2) Primary Neuritic Leprosy:

The cases in which nerve involvement is the result of infection spreading up the cutaneous nerves from a patch of leprosy are known as secondary neuritis. The cases in which the neuritic changes are independent of any existing or past of skin lesions are known as primary or pure neuritis.

Indian classification as suggested by Bhattacharya and Chatterjee (1955) and classification suggested by Wade (1952), has given the name "Pure poly neuritis" for pure neuritis (mono or poly).

### III- ADVERSE EFFECTS OR IMMUNE RESPONSES TO

#### MICROBACTERIUM LEPRAE :

In spite of long generation time of *Mycobacterium leprae* and its low toxicity, leprosy patients may succumb to attacks of acute inflammation in affected tissues which are not due to secondary infection or trauma. Such reactions have been sub classified into numerous types. However, two types of these reactions have been clearly defined

and extensively studied from an immunological point of view during the last decade, namely erythema nodosum leprosum and borderline reactions.

#### (A) Erythema nodosum leprosum (ENL) :

ENL is only found in highly bacilliferous patients, i.e. in BL-LI patients especially when they are put on anti-leprosy treatment. The most common symptom is painful erythematous subcutaneous nodules from which the name ENL is derived (Rea and Levan 1978). The appearance of these nodules is often associated with fever and some times complicated by neuritis, orchitis, tridacyclitis, arthritis, proteusuria, and/or lymphadenopathy. Histology shows microscopic feature skin to Arthus reaction that is perivascular infiltration with granulocytes. Deposition of immunoglobulins and complement (C3) have been shown by immunofluorescence techniques in cases of ENL. However workers have also shown presence of higher levels of C<sub>3d</sub> in plasma of ENL patient (in 70% cases) as compared to those in LI patient (10% cases) (JCMR 1981). Thus in ENL there may be other factors involved in activating complement which are yet to be explored.

Leprosy, like other chronic inflammatory diseases, causes secondary myeloidosis. It is

interesting to note that patients with recurrent EBL reaction seem to especially at higher risk to develop amyloidosis. Other complications reported in lepromatous patients which may be related to circulating immune complexes include glomerulonephritis, poly arthritis and myositis. EBL is an important cause of nerve damage when the reaction takes place in nerve. Another major type of nerve damage in LL appears to be a slow and progressive in nature, primarily affecting cutaneous nerves. Histologically this lesion is characterised by large number of bacilli within Schwann cells, endoneurial and perineurial macrophages and perineurial cells. There is thickening of epineurium with deposition of collagen and fibrosis (Gonal 1978).

#### (B) Borderline reactions :

Borderline patients without bacilliferous lesions may also develop reactions. These lesions are clinically characterised by intracutaneous hyperemia, edema and induration. Such changes may occur in old lesions or new lesions, referred to as lepro-reaction. It is evident that the lepro-reaction is only one type of reaction occurring in the leprosy. It has been assumed that borderline reactions may be subdivided into two types, namely reversal reactions or upgrading reactions and dose grading reactions.

### (1) Reversal reactions or upgrading reactions :

These reactions occur in MT, BB, BL and rarely in LL subpolar type of leprosy. Reversal or upgrading reactions are associated with a movement towards tuberculoid pole. Clinically the lesions are characterised by skin erythema, oedema and peripheral neuritis. Histologically the lesions consist of mainly lymphocytes and epithelioid cells with or without giant cells. Number of bacilli in the lesions are diminished. Lymphocyte transformation test (LTT) and Lymphocyte Migration Inhibition Test (LMIT) to Myco-lepros are generally much stronger than expected. This type of high immunological boost leads to oedema of nerve. Such a rapid upgrading in immunity often leads to nerve damage, consequently deterioration in the form of deformity is noted in such patients.

Serial lymphnode biopsies have been studied in four patients with reversal reactions (Tark and Ester, 1971). Little change was noted in two patients who had mild reversal reactions. However, in two patients who had more severe reactions shifting from LL to BB in one case and MT in other was observed.

### (ii) Down grading reactions :

These reactions are usually mild and occur in untreated MT, BB and BL patients. In such cases, there is paucity of cell mediated immunity.

Consequently in the immunological scale, these patients move down in the spectrum towards the lepromatous pole.

#### **IV- THE NATURE OF IMMUNOLOGICAL DEFICIENCY IN LEPROSY :**

There are two types of immune responses, humoral and cellular. The humoral response is characterized by synthesis of antibody molecule specific to immunising antigen, their release in circulation and hence appearance in the serum. Lack of such detectable serum antibody distinguishes the cellular immune phenomenon. Various *in vivo* and *in vitro* tests have been employed to assess the immunological status of patients.

##### **A- Studies *in vivo* :**

###### **Skin tests:**

Is one the most important diagnostic test for cell mediated immunity is skin testing with appropriate antigen. When the skin tests are evaluated by an experienced observer for the quality (i.e. induration, edema, time course) as well as the size of reaction, they can provide valuable information.

###### **a) Lepromin antigen :**

Studies of immunological reactivity in leprosy patients have demonstrated diverse alteration in the two polar forms of the disease. The lepromatous type of disease goes virtually unchallenged

by the host with his macrophage laden with bacilli, high titre of serum antibodies bathing the tissue, showing negative delayed hypersensitivity to leprosin; and self limited course tuberculoïd type with few lepro bacilli, detectable little or no serum antibody and markedly positive delayed hypersensitivity to leprosin.

Two types of reactions are observed at the site of inoculation of leprosin. A tuberculin like reaction occurring at 24 to 48 hours is called Pernandres (1940) reaction and indurated nodule which appears 2 to 4 weeks later is called the Mitsuda (1919) reaction.

The types of leprosin antigens are commonly used (i) Mitsuda-Hyashi antigen and (ii) Dhamondra antigen. Dhamondra antigen gives a well marked early reaction and weak late reaction whereas Mitsuda-Hyashi leprosin generally gives rise to a weak early and a strong late reaction. The early reaction or Pernandres reaction has been described as a delayed hypersensitivity reaction to soluble constituents of the leprosy bacillus (Dhamondra, 1941) whereas the bacillary component is needed for inducing the late reaction.

Bullock (1960) has compared the response of Mitsuda and Dhamondra antigen in tuberculoïd and

lepromatous patient at 48 hours and at 3 weeks time. He found diminished response in lepromatous patient as compared to tuberculoïd patient with both antigens. Talwar et al (1972), have studied the lepromin test, both early and late reaction, in the spectrum of leprosy and they found that the early as well as late reactions were negative in LL but early reaction was positive in some BL. BB patient showed only positive early reaction while late reaction was negative while BT, TT patient showed both early as well as late reaction as positive. Similar observations have been of other workers (Bodi et al, 1976; Job et al, 1976; Rao et al, 1976; Sharma et al, 1979; Kumar et al, 1980; Rao and Rao 1981).

#### b) Candida antigen :

The difference in the response between tuberculoïd and lepromatous cases of leprosy to inoculation of lepromin has been assumed to reflect a particular type of immune response of patients. Studies have been made to find out the differences in the immune response in leprosy cases using non specific antigen like Candida.

Duck et al (1968) have shown diminished response using *Candida albicans* antigen both in lepromatous as well as in tuberculoïd patients in comparison to controls. Dalloul (1968) studied the

immunological response in tuberculoid and lepromatous leprosy and diminished response was found in both type of cases, but was more diminished in untreated cases than treated cases.

e) Other antigens :

The ability of an individual to respond to various other types of antigens depends upon previous exposure, age, prior testing and other factors.

Sohn and Mittal (1971) have studied normal lymphocyte transfer test (NLT) after intradermal injection of 2.5 million lymphocyte and D<sub>2</sub> Nitrochlore Benzene (DNCB) contact delayed hypersensitivity test in both lepromatous and tuberculoid leprosy. In most of the cases of lepromatous leprosy the NLT reaction has been flat. While in tuberculoid first peak was observed. On the other hand in controls two peaks were observed. This indicates that variable number of cases of both lepromatous and tuberculoid are associated with diminished CMI. The frequency and severity is much more common in lepromatous than tuberculoid. They have also shown that when 100 µgm DNCB was challenged to the patients, only 10% of lepromatous and 16.6% of tuberculoid patients gave the positive results. On the other hand when the patients were challenged with 400 µgm, 60% lepromatous cases and

100% tuberculoid cases showed positive results. When dose was increased up to 1000 ugs then 77% lepromatous and 100% tuberculoid cases showed positive results. Thus if a weak stimulus has not been sufficient to attract the necessary number of healthy immunocompetent cells out of total number of lymphocyte population crowded at the test site, then in the same case a stronger stimulus may attract the required number of immunologically competent cells necessary for expression of delayed allergic response explaining thereby why a bigger dose of an antigen could produce a positive skin test in cases in which lesser dose of same antigen has failed to induce it. Similar observations have been of other workers (Guinto 1962; Leifer 1969; York and Waters 1969).

Ban et al (1974) have studied 42 lepromatous leprosy cases using streptokinase, streptodornase, mumps antigen, trichophyton and histoplasma. They found diminished delayed hypersensitivity response in lepromatous leprosy cases as compared to controls.

Recently Kumar et al (1980) studied skin delayed hypersensitivity test using PPD, DNBC, Concanadine, Croton oil and histostim. They have found that lepromatous group show either very mild

reaction or negative reaction. While significant response in BB and TT was observed. Thus showing the importance of cutaneous skin test in BB to detect the shift towards any end of the immunological spectrum.

#### B- Studies in vitro :

##### 1) Status of T and B-cells :

Barry et al (1948) had shown that the lymphocytes were involved in immunological mechanism. It is now recognized that lymphocytes form an indispensable component of bodies immune system and embryonic precursor of cells that will give rise to both cell mediated and humoral immune responses.

Studies by Glaman et al (1966), Davis et al (1967); Miller and Michell (1968) indicated that at least two population of lymphocytes were involved in most of immune responses and they differed in their anatomical distribution. Cells of one population were the precursor of plasma cells. They were present in the bone marrow but not in the thymus and they corresponded to the bone dependent system of chickens. Another population was dependent on and are derived from thymus and although they had an obligatory role in most antibody responses, they did not themselves turned into plasma cells. These two population of lymphocytes are currently known as T-cell (Thymus dependent) and B-cell (Bone equivalent derived) (Miller et al 1969).

Groves et al (1973) had shown that T-cells appear to be concerned with cell mediated immunity and B-cells with humoral immunity. T-lymphocytes play a major part in immune response to facultative organism, tissue or organ graft and certain infection with viruses, B-lymphocytes mature to become antibody producing plasma cells and play a role in humoral antibody responses (Routledge 1975).

T and B lymphocyte population comprise a number of functionally different subsets and it may be possible to distinguish between these two subsets. T-cells are recognized by their ability to bind with sheep red blood cells spontaneously in a characteristic morphological configuration termed as rosette (Pulsenberg, 1973). While human B-cells possess surface immunoglobulin detectable by direct immunofluorescence (Gelmann, 1974). They also possess receptors for aggregated immunoglobulins, for antigen antibody complex and for third component of complement (C3). These receptors are detectable by erythrocytes coated with antibody and complement that surround B-lymphocytes in a cluster (Mendes et al 1973).

Immunity to intra-cellular organism is dependent on cell mediated immune mechanisms rather than humoral antibodies. Further more studies on

experimental animals have revealed that carriers of this immunity are T-cells. However T-cells are capable of killing the organism directly but this function is accomplished through the molecular phagocytes (macrophages). At least two phases seem to be involved. In initial phase when foreign antigens are encountered in tissue, T-cell increases the antibacterial activity of surrounding macrophages and this is called as "macrophage activation phase". These macrophages are stimulated by the liberation of lymphokines (molecular mediator) from the T-cells. Later on chemoatetic substances are released which increase the influx of macrophage precursor (monocyte) in lesion. This phenomenon has been called as macrophage mobilization (Godal, 1978).

From above description, it is evident that T-cell status is an important factor in the assessment of cell mediated immunity. Various workers have shown that the T-cell count in the peripheral blood had decreased gradually in the spectrum of leprosy from tuberculoid pole to lepromatous pole; maximum fall being in lepromatous and minimum being in tuberculoid pole. Contrary to this T-cell count is found to be increased towards lepromatous pole than tuberculoid pole (Purohit et al 1973; Gajapeewalka et al 1973; Liu et al 1974; Bhagat et al 1977; Sharma et al, 1979; Sachdev et al, 1980). Verma et al (1971) had

studied only B-cell status obtained by teasing the lymphnode from lepromatous leprosy patients and found that B-cell count was increased in the lymphnode of lepromatous leprosy cases as compared to B-cell count from lymphnode of normal human being. It is similar to the findings of B-cell count in peripheral blood by other workers.

Mendes et al (1974) have studied T and B cells in peripheral blood as well as in lymphnode of lepromatous leprosy cases. A significant decrease in proportion of T-cells was observed in peripheral blood and depletion of T-cells in percutaneous areas of involved lymphnode indicating impaired cell mediated immunity. B-cells were found to be increased in peripheral blood as well as preservation of B-cell area was seen in lymphnodes. Similar observations have been obtained by Methias et al (1980) in the study of cellular changes in spleen.

#### ii) Status of <sup>p</sup>suppressor cells :

It has been established that there is failure of T-cells in lepromatous patients to respond to antigens of leprosy bacillus. The mechanism for this selective unresponsiveness remains unknown (Godal 1978).

It has been postulated that unresponsiveness of lepromatous patients to the antigen of leprobacillus, and their possible responsiveness to related antigen

of tubercle bacillus would be due to presence of a specific population of suppressor lymphocytes capable of being triggered by at least one unique antigen of leprosy bacillus. Mehra et al (1980) in their study have shown the ability of lymphocytes from leprosy patients exposed to Dhamendra lepromin to suppress the response of the population of cells to the T cell mitogen, Con A. Significant Myco-lepro induced suppression of Con A response was found with peripheral blood lymphocytes of 22 out of 35 lepromatous patients and 18 out of 18 patients with borderline lepromatous or borderline tuberculoid. In contrast, lepromin-induced suppression of only 2 out of 11 tuberculoid patients and 2 out of 30 normal donors was observed, indicating a correlation between suppression in vitro and the degree of unresponsiveness observed in the patients.

### iii) Status of macrophages :

The macrophage, a phagocytic cell endowed with bactericidal power in their lysosome, plays a vital defensive role in microbial invasion in host. In diseases such as leprosy, which are caused by bacteria growing intra cellularly and mainly inside macrophages, this cell is undoubtedly the immediate effector cell which is responsible for death and

elimination of the pathogenic agent. The elegant studies of Neckameier (1969) clearly suggest that the bactericidal and bactericolytic properties of macrophages in this type of infection reach their fullest expression of these cells by specifically committed lymphocytes.

In 1967, Barbieri and Correa reported that macrophage from Mitsuda-negative individuals were inactive *in vitro* against *Mycobacterium leprae*, while macrophages from Mitsuda positive persons caused the lysis of bacillus *in vitro*. Similar results have been observed by Beigneux (1967) and Piscani et al (1973).

Recently Bindi et al (1980) have shown that macrophage from lepromatous patients after phagocytosis of *Mycobacterium leprae* showed alteration in the surface property as determined by their ability to express Fc receptor. The same macrophage without intracellular *Mycobacterium leprae* show normal Fc receptor. The lepromatous macrophages also show very poor interaction with lymphocytes in presence of *Mycobacterium leprae*, while they are able to interact with lymphocytes when exposed to other antigens. There appears to be a defective macrophage population in lepromatous patients that is unable to process *Mycobacterium leprae* antigens and initiate the CMI response.

iv) Lymphocyte blast transformation test :

Mitogenic response of peripheral lymphocyte to phytohaemagglutinin (PHA), in vitro has been used to assess the functional capacity of T-cells, which is an indirect assessment of cell mediated immune response.

Pierka and Shepard (1968) have studied the blastogenic response in leprosy patients using PHA, PPD, and BCG in a 5 days culture of peripheral lymphocyte at 37°C and found that most of lepromatous leprosy cases had markedly depressed lymphocyte response to PHA as well as to mycobacterial antigens. The response to PHA was only moderately depressed in patients with tuberculoid leprosy. Similar type of result had been also observed by other workers either using only PHA or other antigens for 3 to 7 day. (Rao et al, 1971; Puri et al 1971; Nelson et al 1971; Godal et al 1971; Bullock et al 1971; Nehru et al 1972; Ulrich et al 1972; Talwar et al 1972; Lin et al 1973; and Job et al 1976).

Kahlonanis et al (1977) had studied the PHA response alongwith T-lymphocytes in peripheral blood and had suggested that depressed response to PHA was associated with reduction in circulating T-lymphocytes. The other workers had also observed the same response (Rao et al 1976; Nath et al 1977; Sharma et al 1979; Ghai et al 1980; Dubey et al 1981).

v) Leucocyte migration inhibition test (LMT) :

LMT appears at present to be the most promising test for evaluation of cell mediated immunity. Myrvang et al (1973) have studied the OMI response using lymphocyte transformation test as well as LMT and found diminished response from TT to LL. Similar have been the observations of Rao and Rao (1981).

vi) Serology of leprosy :

There are evidences that antimycobacterial antibodies are produced in leprosy and these circulating antibodies probably do play an important role in the immunopathology of lepromatous leprosy. Watanabe et al (1969) have reported the presence of immunoglobulins, complement and soluble mycobacterial antigens in the lesions of erythema nodosum leprosum (ENL).

Noronha et al (1973); Rojas-Laplaque et al (1972) and Gelber et al (1974) have demonstrated substances in the sera of patients with ENL which precipitate with C<sup>1q</sup> component of complement system and may be immune complexes. Several studies have shown the presence of renal lesions, particularly in patients with ENL, which

are consistent with pathology of immune complex glomerulonephritis (Shoe, 1972; Bruts and Gutman, 1973; and Bullock et al, 1974).

Autoantibodies of many type, including rheumatoid factors, anti-thyroglobulin precipitins, antinuclear antibodies and antibodies which fix to intercellular areas of epithelial surfaces have been reported in lepromatous patients. In vivo fixation of immunoglobulin in basement membrane zone of skin of lepromatous patients has been reported by Bullock et al (1974) and Quiñones et al (1975).

Various serological tests are also carried out for early diagnosis of leprosy. These include leproagglutination test, indirect fluorescent antibody test (IFA-ABS) and Radio-immuno assay.

\*\*\*

## **MATERIAL & METHODS**

The study has been conducted on patients, suffering from various types of leprosy, admitted to the Department of Skin & V.D., M.L.D. Medical College hospital, Jhansi, between June '81 to March '82. Age and sex matched patients with minor ailments without immunological disorders were selected as controls.

The patients were thoroughly examined clinically for type of leprosy and findings were recorded on the pre-designed proforma (Appendix I).

10 ml heparinized peripheral blood samples (25 unit heparin per ml) were collected in sterile tubes from patients, under all sterile conditions. Blood was simultaneously collected from all these patients for routine haematological tests.

Nozal and slit skin smears were prepared from each patient.

#### METHOD FOR PREPARATION OF SLIT SKIN SMEAR :

It was most important to choose right part of skin from which to make the smears. The edge of an active lesion i.e. area which was reddened, thickened or raised, or free middle of lesion, when it was flat, was selected for slit skin smear.

The part was cleaned with the spirit swab and then the area was pinched between thumb and index finger. A small shallow approximately 5 mm long cut was made with sterilized scalpel blade and then

margins and bottom were scraped with edge of blade and scrapings were smeared on the already flamed and cleaned slide. The smear was fixed with flame of spirit lamp. It was stained with modified Ziehl-Neelsen's method using boiled concentrated carbolic fuchsin for 10 minutes and decolourised with 1% acid alcohol and counter stained with 1% methylene blue and observed under oil immersion objective.

#### NASAL SWAB :

The patient was asked to sit on a stool facing towards good light. A speculum was put in the patient's nose. The nasal was flamed and cooled. The nasal septum was scraped gently with the help of the spud and then the scrapings were smeared on a cleaned and flamed slide. It was fixed by gentle flaming and stained as skin slit smear. Bacterial assessment was done according to Ridley and Jopling (1966) as follows:-

- '0' No bacilli in 100 oil immersion fields.
- +' 1-10 bacilli on average in 100 oil immersion fields.
- ++ 1-10 bacilli on average in 10 oil immersion fields.
- +++ 1-10 bacilli on average in one oil immersion field.
- ++++ 10-100 bacilli average in one oil immersion field.
- +++++ 100-1000 bacilli on average in one oil immersion field.
- ++++++ Many clumps or globi in one oil immersion field.

### SKIN BIOPSY :

Skin biopsy was taken from the active edge of the lesion and fixed in 10% formaline.

The biopsy was processed in concentrations of alcohol, then cleaned with Xylene in place of chloroform. Tissues were embedded in paraffin (55°-60°C); blocks prepared and 5  $\mu$  thick sections were cut with the help of rotary microtome. In each case, one slide was stained with haematoxyline and eosin stain and other section was stained with modified technique of Pite-Parese staining for demonstration of *Leprosy bacilli* (Gullings 1974).

### HISTOPATHOLOGICAL TYPING :

The histopathological typing of cases were done according to criteria of Ridley and Jopling (1966) as given below :-

#### Tuberculoid type (T.T.) :

- Erosion of basal layer by granuloma.
- Pool of well developed epitheloid cell granuloma with few Langhan's giant cells often enveloped by dense zone of lymphocyte, especially in deeper part of dermis.
- Nerves difficult to detect and may show coagulation.
- *Leprosy bacilli* were not detected.

### **Border line tubercleoid ( B T ) :**

- Narrow clear sub epidermal zone above the granuloma.
- Lymphocyte plentiful and diffuse.
- Nerves swollen but recognizable.
- Lepra bacilli demonstrable 0 to ++.

### **Border line ( B B ) :**

- Sheets of epithelioid cells but no giant cells.
- Lymphocyte sparse and diffuse.
- Nerves showing structural disorganization, but no granuloma.
- Lepra bacilli demonstrable in grade +++ to +++++.

### **Border line lepromatous ( B L ) :**

- Granuloma with cells slightly epithelioid in appearance.
- Few lymphocytes.
- Lepra bacilli demonstrable in grade +++++ to ++++++.

### **Lepromatous leprosy ( L L ) :**

- Macrophages with some foamy change heavily laden with bacilli.
- Very scanty lymphocytes.
- Nerves almost normal.
- In active phase, lepra bacilli +++++ to ++++++.

or

- In regressing phase, form cells with globi and much fat. A few lymphocyte, lepra bacilli ++++ to +++++.

#### TOTAL LEUCOCYTE COUNT ( T . L . C ) :

The total and differential leucocyte count was done by techniques described by Dacie and Lewis (1975).

#### ABSOLUTE LYMPHOCTYE COUNT ( A . L . C ) :

The absolute lymphocytes were calculated in every case from total and differential leucocyte counts using the following formula.

$$A . L . C = \frac{T . L . C \times \% \text{ of lymphocyte}}{100}$$

#### EVALUATION OF T AND B LYMPHOCYTES AND P. H. A. RESPONSE :

T and B lymphocytes present in peripheral blood were demonstrated by means of their surface receptors (Jondal et al 1972).

The basic principle of procedures is as below :

- Separation of lymphocytes.
- Demonstration of T cell by Sheep Red blood cell (SRBC) rosette (Rosette).
- Demonstration of B cell by formation of rosettes with SRBC coated with ant sheep haemolysin antibody and complement.

### Lymphocyte separation :-

The lymphocytes were separated by Ficoll Conray 420 density gradient centrifugation. The differences in density for the various cells in blood make it possible for them to be separated by this method.

### Material required :-

- 1- Ficoll Conray 420 solution specific gravity 1.077 was prepared (See Gupta, 1981). (11.40 gm. of Ficoll dissolved in 160 ml of distilled water + 22 ml of Conray 420. Specific gravity was adjusted to 1.077 and sterilized by Soitz filter).
- 2- Heparin - preservative free.
- 3- Minimum essential medium ( Eagle ) with Hanks base (Micro Lab Bombay ).
- 4- TC Medium 199 ( Difco Laboratories, Detroit Michigan, U S A ).
- 5- PHA-M ( Difco Laboratories, Detroit Michigan, U S A ).
- 6- Almevera solution :-

Glucose	...	...	24.6 gm
Trisodium citrate (dehydrate)	...	...	9.6 gm
NaCl	...	...	30.04 gm
Distilled water	...	...	1200 ml

pH was adjusted to 6.1 with 10% citric acid.  
Sterilized by low pressure autoclaving.

**7- Phosphate buffer saline ( PBS ) :**

- Phosphate buffer solution :

(A) 0.15 M - Na<sub>2</sub>HPO<sub>4</sub>, NH<sub>4</sub>O 22.4 gm/litre

(B) 0.15 M - Na<sub>2</sub>HPO<sub>4</sub> 21.3 gm/litre

**Normal saline**

Na Cl 9.0 gm/litre

- Phosphate buffer saline

For pH 7.4 - Mix solution 'A' - 18 ml

'B' - 62 ml

and then Normal saline 100 ml was added.

Solution was sterilized by autoclaving.

**8- Glutaraldehyde - 2% solution in PBS.**

**9- A B sera collected from persons of A B blood group.**

**10- Complement from mouse.**

**Method :**

- 2.5 ml Picoll solution was taken in two sterilized test tubes.
- 5 ml of blood was layered over each test tube carefully from sides of test tubes and centrifuged immediately at 1500 rpm for 15-20 minutes.
- Various layers formed after centrifugation contain plasma, predominantly lymphocyte, Picoll沉降, neutrophil and red cells.

- After removing plasma carefully the lymphocyte layer was pipetted off and placed in 5 ml of MEM from one test tube and 5 ml of TC medium 199, and washed twice, centrifuging each time at 1000 rpm for 10 minutes with MEM and TC medium 199 respectively.
- The cells were re-suspended in MEM and viability was checked by 1% eosin TC medium 199 and count was adjusted to  $2-3 \times 10^6$  cells/ml T-cells (SRBC Rosette for T-cells) :
  1. Sheep red blood cells (SRBC) were collected in Alsever's solution and washed thrice in phosphate buffer saline (PBS) and suspension made up to 0.5% in PBS.
  2. 0.5 ml of lymphocyte suspension was mixed with 0.5 ml of SRBC and incubated for 15 minutes at 37°C.
  3. Then incubated at 4°C over-night.
  4. 1 ml of 2% Glutaraldehyde was added and then test tube was kept in ice for 15 minutes.
  5. Finally the wet preparation was made and stained with 0.5% methylene blue and 200 cells were counted.

**Result :**

Three or more SRBC adhering to lymphocyte were taken as rosette forming cells. The absolute T-cell count was calculated as below :

$$\text{Absolute T-cell count} = \frac{A10 \times \% \text{ of T-cell}}{100}$$

**B-cells (EAC Rosette) /for B-cells :**

1. SRBC were washed thrice in PBS and adjusted to a concentration of 5%.
2. To 0.5 ml of 5% SRBC was added and incubated for 15 minutes at 37°C with 0.5 ml of antisheep haemolysin (Amboceptor). Subhaemolytic dose of Amboceptor was assessed before putting the the test (Crichtonshank 1975).
3. SRBC were washed thrice with PBS and resuspended with 0.5 ml of PBS.
4. 0.5 ml of 1:10 dilution of complement (mouse) was added to SRBC and incubated for 45 minutes at 37°C.
5. These SRBC (i.e. EAC now) were washed thrice with PBS and resuspended to make 0.5% concentration in PBS.
6. 0.5 ml of lymphocytes were added to 0.5 ml of 0.5% of EAC in PBS and incubated at 37°C for 30 minutes.
7. The solution was resuspended and wet preparation was prepared and stained with 0.05 methylene blue and then 200 cells were counted.

**Result :**

Three or more SRBC adhered to lymphocyte were considered to be rosette. Absolute B-cell count was calculated as below :

$$\text{Absolute B-cell count} = \frac{\text{ALC} \times \% \text{ B-cell}}{100}$$

**Lymphocyte blast transformation test :**

When lymphocytes are exposed to antigen against which they are sensitized, they respond by enlargement and cell division. This phenomenon has been called as lymphocyte blast transformation. Lymphocyte blast transformation test was done by using PHA-M as antigen. Technique of Godal et al (1971) was adopted.

Cultures were set as follows :

	Control	Experiment
Lymphocyte suspension in TC medium 199.	0.5 ml	0.5 ml
AB sera	0.5 ml	0.5 ml
TC medium 199	1.0 ml	0.90 ml
PHA-M	nil	0.02 ml

- all cultures were kept in duplicate and were incubated at 37°C for 72 hours.
- Harvested and washed thrice with TC medium 199 and cells were suspended in 0.75 ml TC medium 199.

- Smear was made, fixed in methanol for 5 minutes, stained with Leishman's stain and seen under oil immersion of light microscope.
- 200 Cells were counted and percentage of blast cells were calculated.

#### DELAYED HYPERSENSITIVITY TEST :

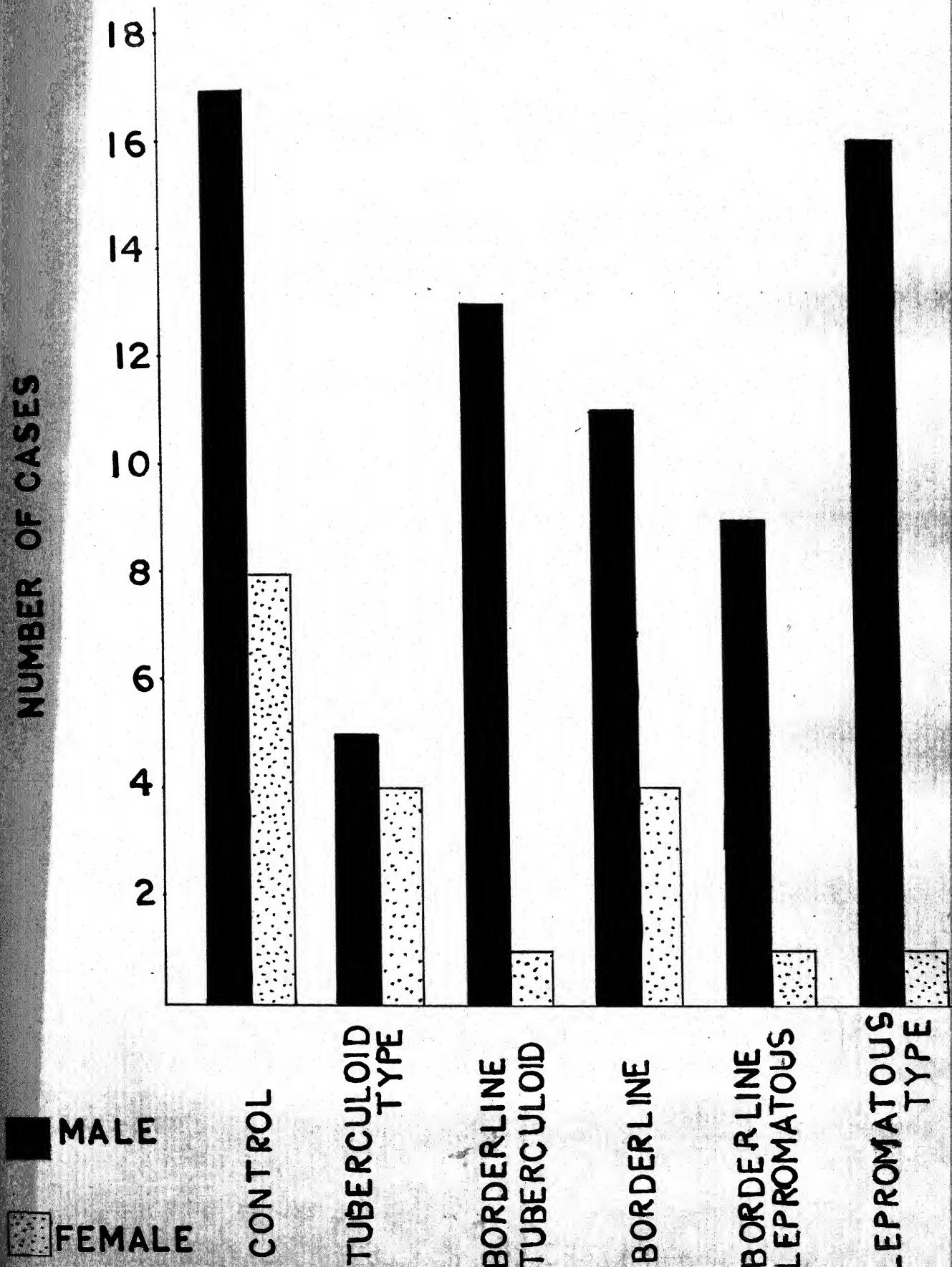
##### Lepromin test :

0.1 ml of Pharsadra antigen, prepared in Central JADIA Institute for leprosy, Taj Ganj, Agra, was injected intradermally in flexor aspect of left arm of patients and control cases. The early (Pernandez) reaction was read 48 hours after the injection and late (Mitanda) reaction was read after 3 weeks. For early (Pernandez) reaction 10 mm or more zone of induration was taken as positive (Guitto 1968). Recommendation of VII International Congress of Leprosy (1959) was followed and 5 mm or more of induration was taken as positive for late (Mitanda) reaction.

##### Delayed hypersensitivity test using Canilide antigen :

0.1 ml Canilide antigen (Prepared in CSIR Centre for Biochemistry, V.P. Chest Institute Building, Delhi) was injected intradermally in right fore-arm of patients as well as in control cases. The test was read after 48 hours and induration of 5 mm or more was taken as positive (Buck et al 1968).

# SEX DISTRIBUTION OF CONTROL CASES & DIFFERENT TYPES OF LEPROSY CASES.



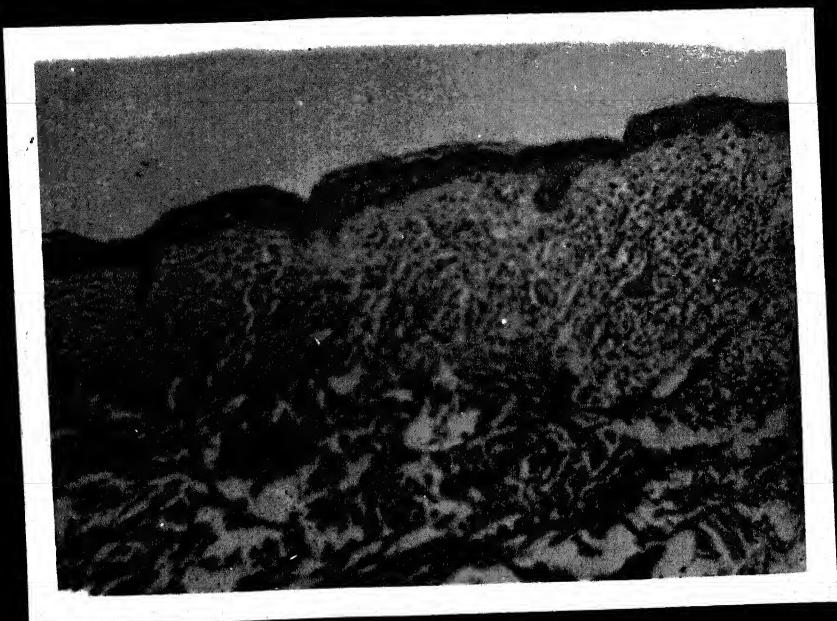


Fig.2a: Skin showing tuberculoid type of  
Leprosy(10 x 7)(H & E staining)

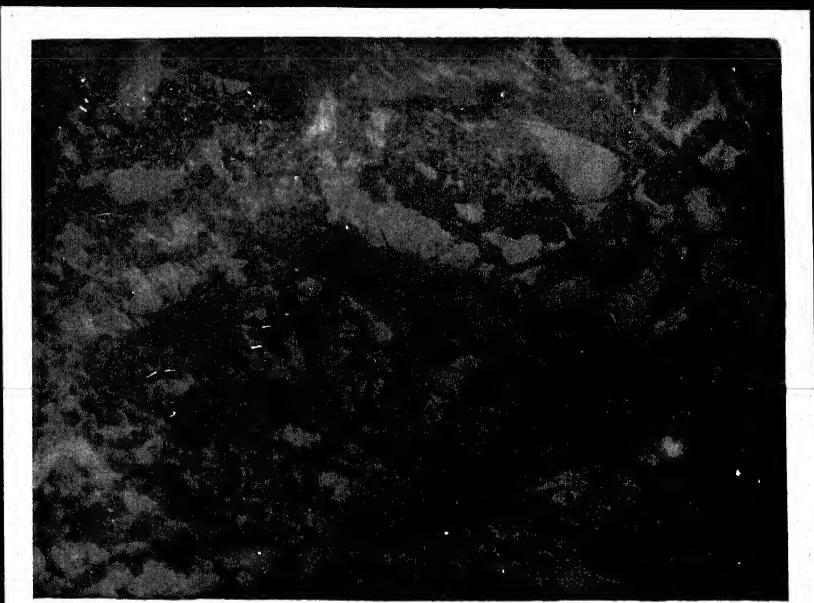
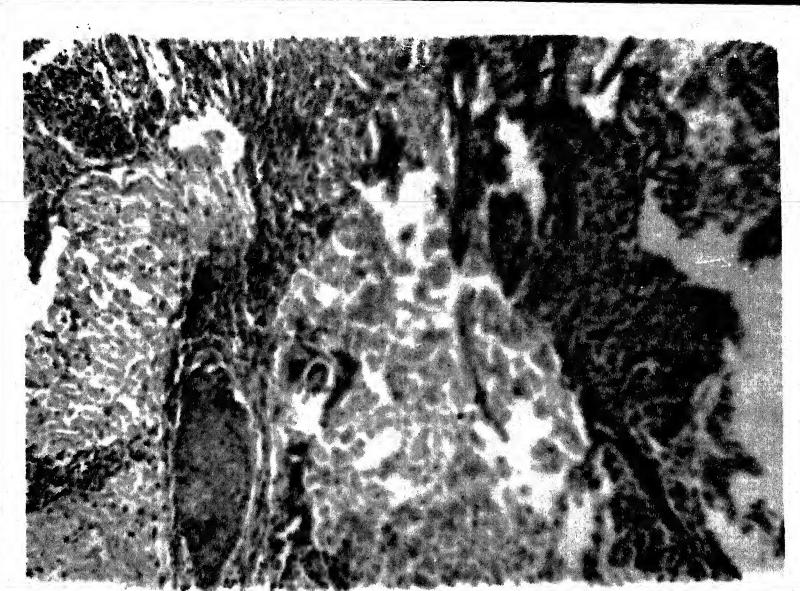
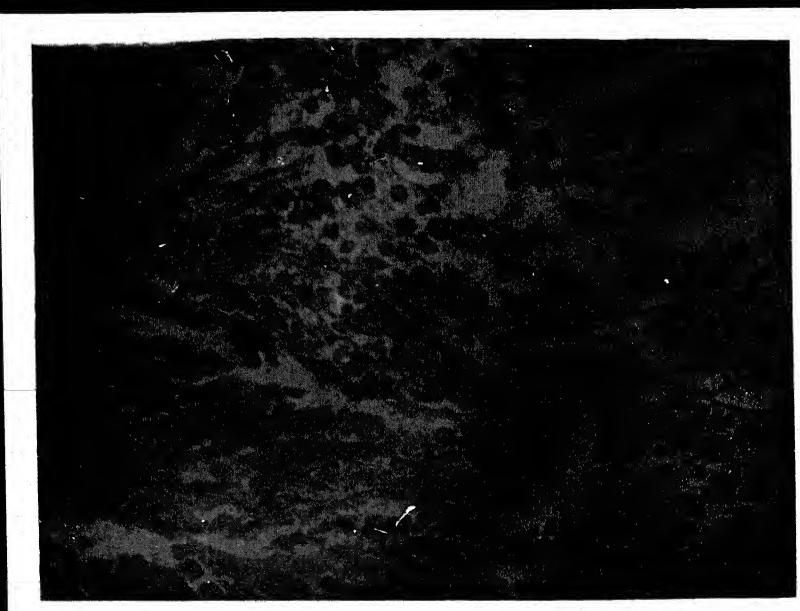


Fig.2b: Skin showing tuberculoid type of  
Leprosy(10 x 7)(H & E staining)



**Fig.3a:** Skin biopsy showing borderline tuberculoid type of Leprosy (10 x 7) (H & E staining).



**Fig.3b:** Skin biopsy showing borderline tuberculoid type of Leprosy (40 x 7) (H & E staining).



Fig.4 a: Skin biopsy showing borderline type  
of leprosy (10 x 7) (H & E staining)

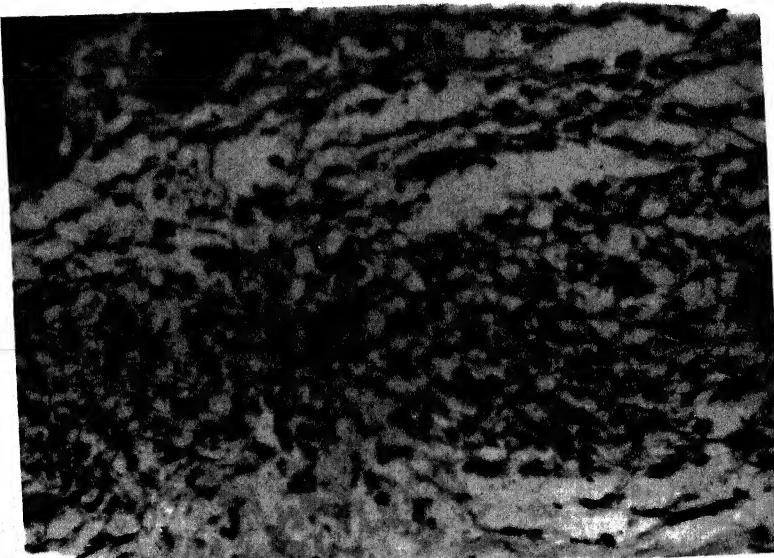


Fig.4b: Skin biopsy showing borderline type  
of leprosy (40 x 7) (H & E staining)



Fig. 6a: Skin biopsy showing lepromatous  
Leprosy (10 x 7) (H & E staining)

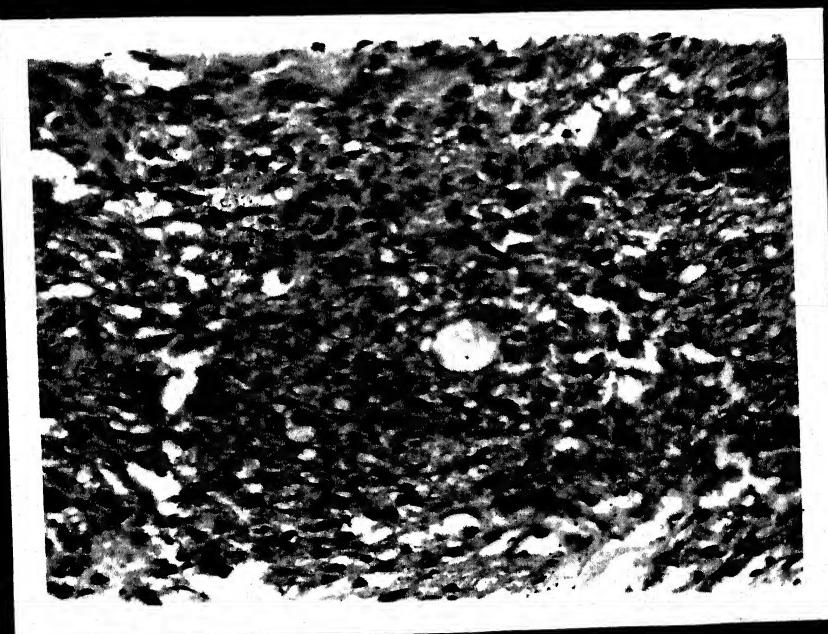


Fig 6b.: Skin biopsy showing lepromatous  
leprosy (40 x 7) (H & E staining)

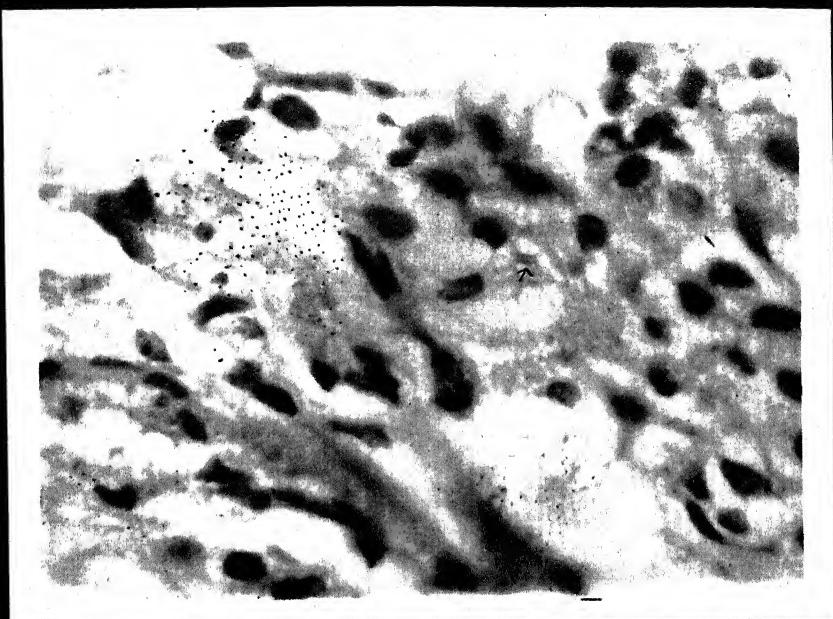


Fig.7a: Skin biopsy showing *Mycobacterium leprae* (isolated) (modified Zeithl Neelson) (100 x 7) (H & E staining)

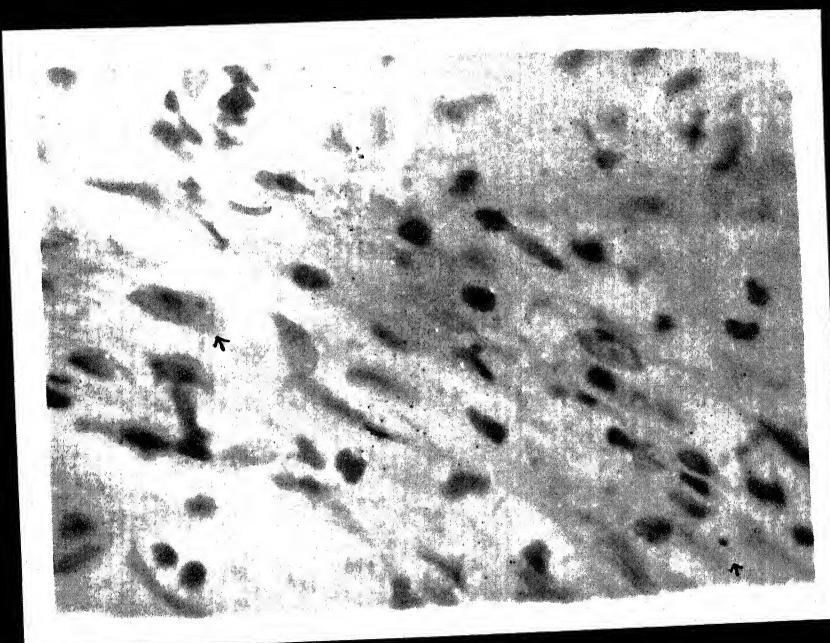


Fig.7b: Skin biopsy showing *Mycobacterium leprae* (globi) (modified Zeithl Neelson) (100 x 7) (H & E staining)

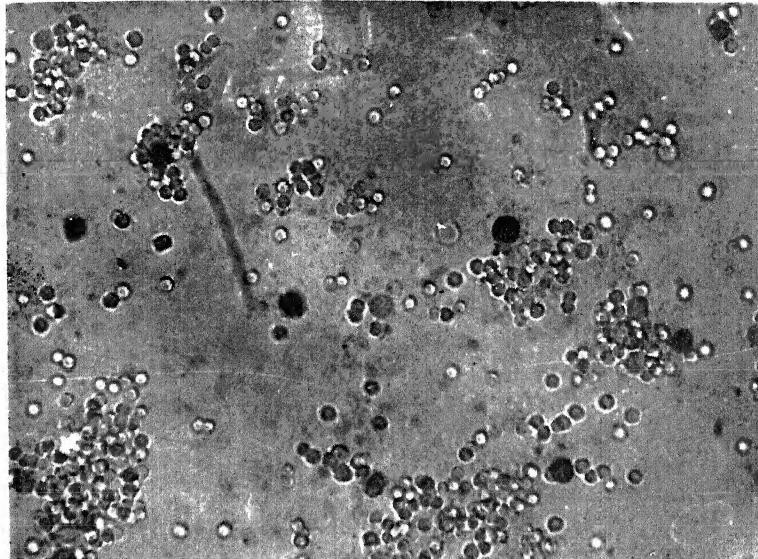


Fig.8: T-cell rosette (E-rosette) (40 x 7).

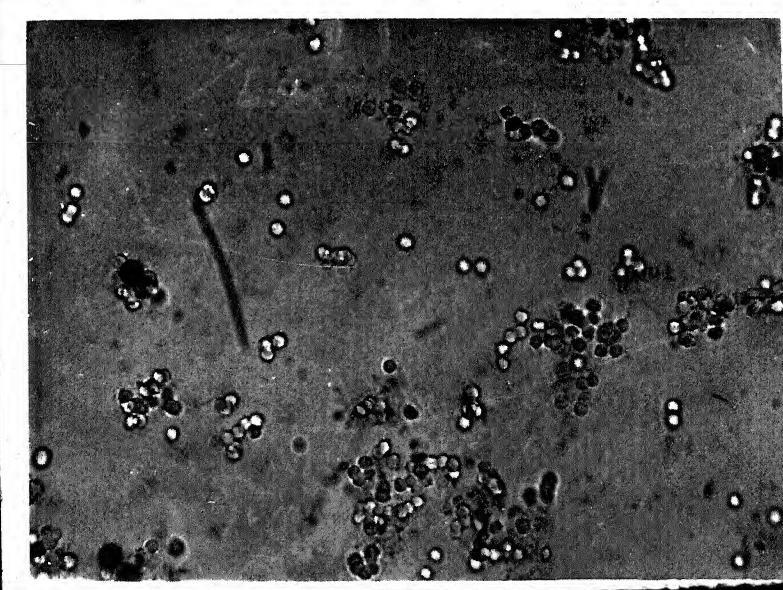


Fig.10: B-cell rosette (RBC rosette) (40 x 7).

# T- CELL % IN CONTROL CASES & DIFFERENT TYPES OF LEPROSY CASES.

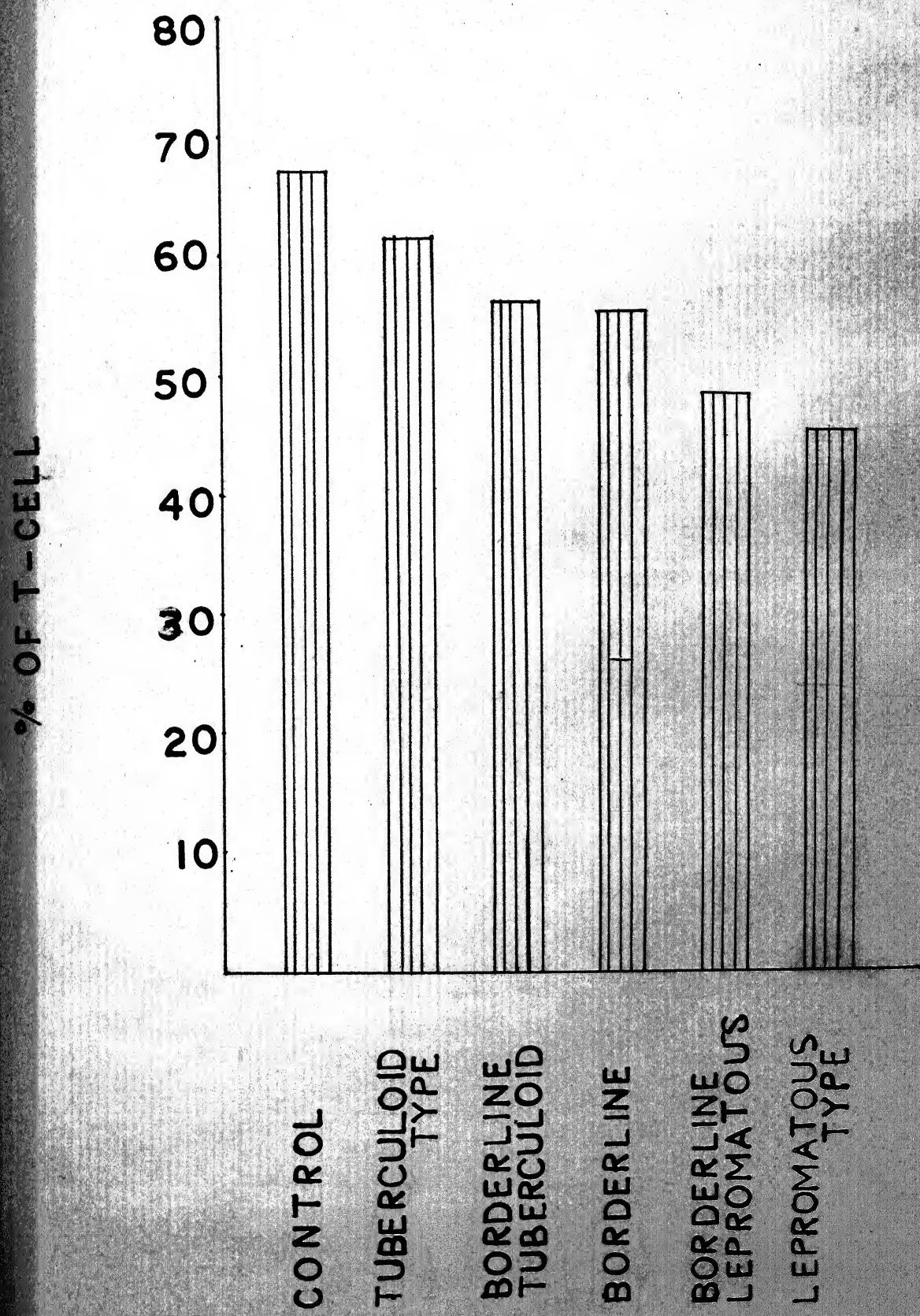
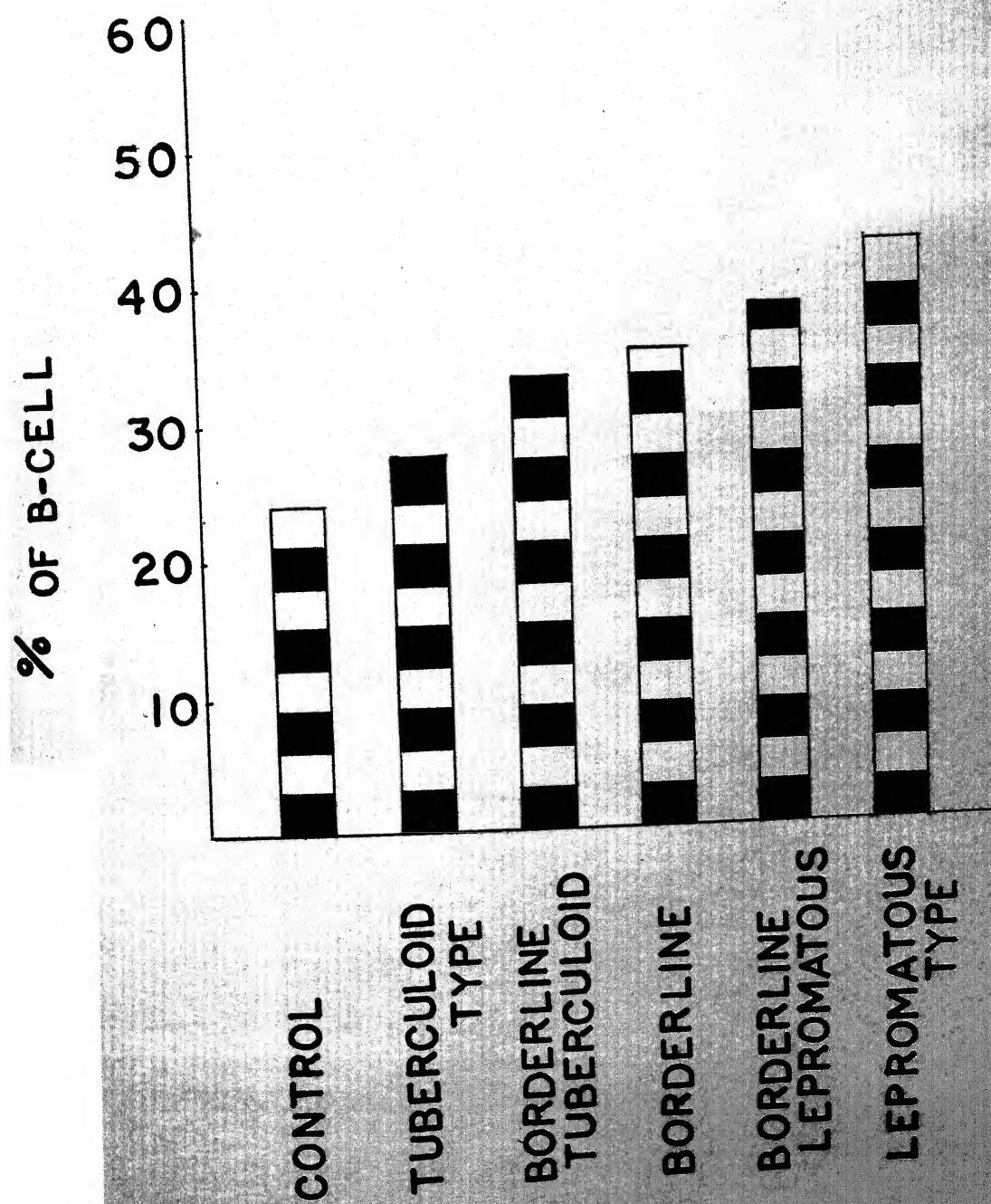


FIG. 9

B - CELL % IN CONTROL CASES & DIFFERENT  
TYPES OF LEPROSY CASES.



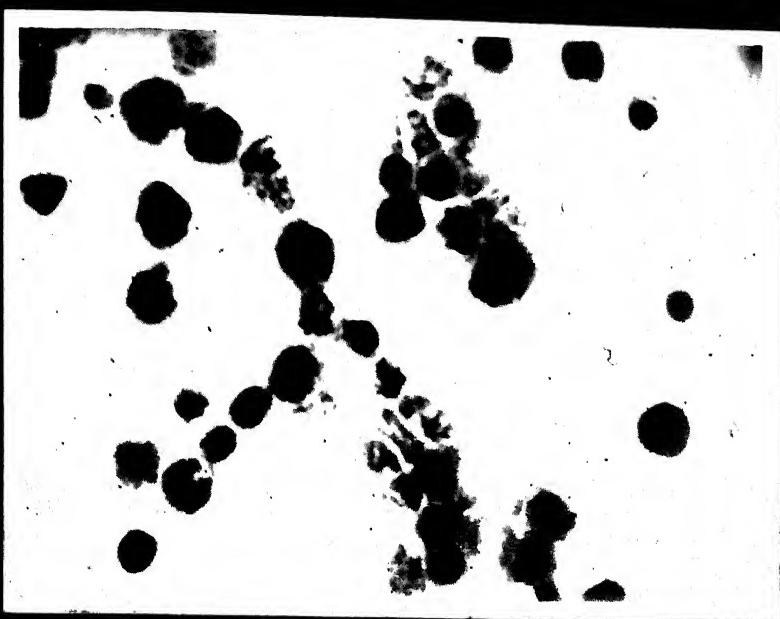
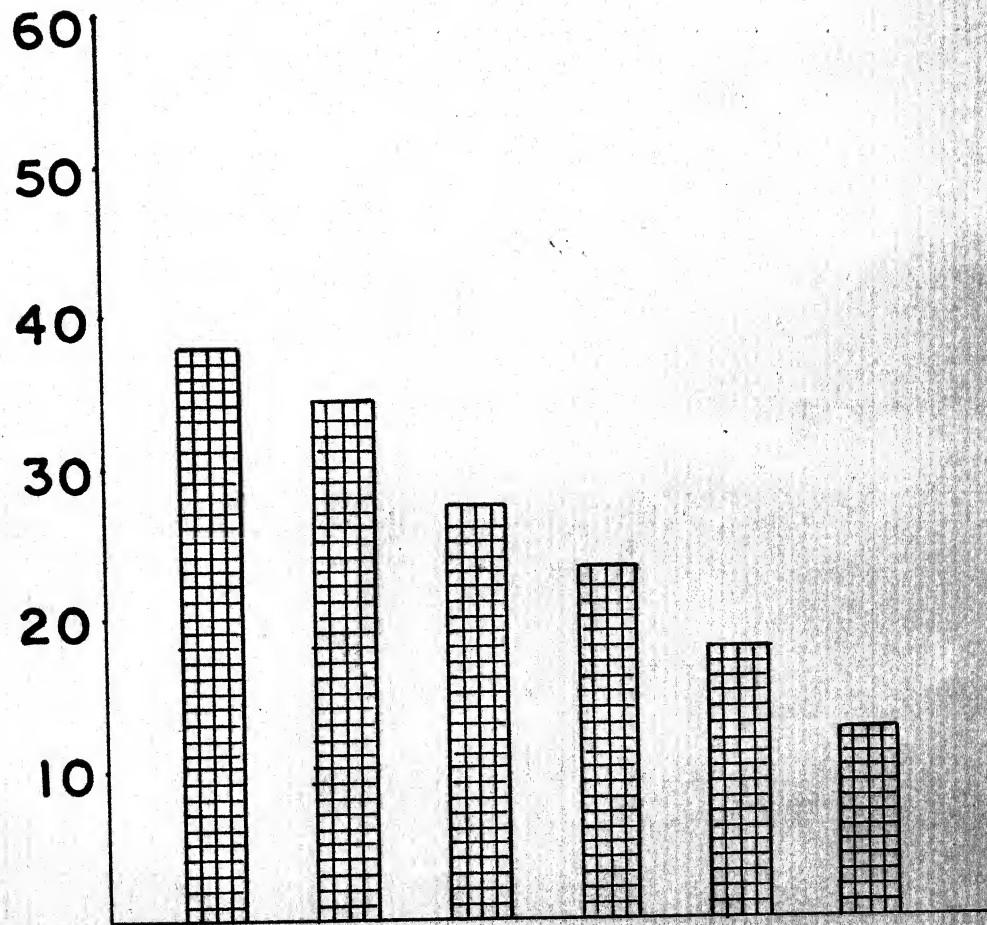


Fig.12 : Lymphoblast transformation after  
PHA response.

# LYMPHOBLAST TRANSFORMATION % IN CONTROL

CASES & DIFFERENT TYPES OF LEPROSY CASES.

% OF LYMPHOBLAST TRANSFORMATION



CONTROL

TUBERCULOID  
TYPE

BORDERLINE  
TUBERCULOID

BORDERLINE

BORDERLINE  
LEPROMATOUS

LEPROMATOUS  
TYPE

FIG. 13

# OBSERVATIONS



The present study was carried out on 90 cases out of which 65 were of various types of leprosy and 25 were age and sex matched controls (Table I).

**Table I**  
**DISTRIBUTION OF CASES**

Groups	Number of cases
Control cases	25
Study group cases	65
Total	90

The control group included relatives of patients admitted to this hospital, junior doctors of the institution and patients suffering from minor ailment like hydrocoele and varicella who were not under treatment of any drugs known to affect immunological status of host.

#### **Study Group Cases:**

The distribution of 65 patients included in the study according to histology is shown in Table II. Out of 65 cases, 9 were of tuberculoid type (T), 46 were of borderline tuberculoid (BT),

18 were of tuberculous (TB), 20 were of borderline lepromatous (BL) and 17 were of lepromatous type (LL).

Table II

**DISTRIBUTION OF CASES ON THE BASIS OF HISTOPATHOLOGICAL CLASSIFICATION.**

Type of cases	Number of cases	Percentage
Tubercloid type	9	13.84
Borderline tubercloid	16	21.53
Border type	18	26.07
Borderline lepromatous	10	15.41
Lepromatous type	17	26.15
<b>Total</b>	<b>65</b>	<b>100.00</b>

Age and sex distribution of control and study group cases is shown in Table III. In study group, age varied between 10 to 60 years with mean ( $\pm SD$ ) of 38.27 ( $\pm 10.68$ ) and in control group between 14 to 58 years with mean ( $\pm SD$ ) of 38.5 ( $\pm 9.99$ ). Out of 65 cases in study group, 56 were male and 9 were female and in control group, 37 were male and 8 were female.

The mean age ( $\pm$  SD) of male and female in control group was 35.5 ( $\pm$  9.70) and 39.35 ( $\pm$  8.56) respectively and in study group was 37.35 ( $\pm$  15.06) and 40.77 ( $\pm$  20.57) respectively.

Table III  
AGE AND SEX DISTRIBUTION OF CASES

Age range Years	Control group			Study group		
	Male	Female	Total	Male	Female	Total
11 - 20	-	1	1	6	-	6
21 - 30	7	4	11	11	8	19
31 - 40	6	3	9	10	3	13
41 - 50	5	1	6	10	4	14
51 - 60	2	-	2	7	-	7
61 - 70	-	-	-	2	2	4
<b>Total</b>	<b>27</b>	<b>9</b>	<b>36</b>	<b>56</b>	<b>11</b>	<b>68</b>
Mean	35.5	39.35	38.5	37.35	40.77	38.57
SD	9.70	8.56	9.99	15.06	15.05	15.60

Age distribution in different types of leprosy cases is shown in Table IV. The majority of cases were within third to fifth decade of life. The mean age ( $\pm$  SD) of cases in TT, BT, BB, BL and LL was 35.5 ( $\pm$  9.42), 35.5 ( $\pm$  13.68), 37.5 ( $\pm$  13.45), 38.5 ( $\pm$  21.50) and 41.30 ( $\pm$  21.91) respectively.

**Table IV**  
**AGE DISTRIBUTION OF DIFFERENT TYPES OF CASES.**

Age Groups Years	TT	WT	WD	WL	WL	Total
21 - 30	-	3	-	3	1	6
31 - 40	3	3	3	1	3	13
41 - 50	4	3	3	3	4	18
51 - 60	3	4	3	2	7	19
51 - 60	2	2	1	1	2	7
61 - 70	-	-	1	3	1	5
<b>Total</b>	<b>9</b>	<b>16</b>	<b>15</b>	<b>10</b>	<b>17</b>	<b>66</b>
Mean	22.5	22.5	27.5	22.5	21.50	28.27
$\pm$	5.45	5.45	5.45	5.30	5.94	10.68

Sex distribution of different types of cases is shown in Table V (Fig.1). Out of 9 cases in TT group 8 were male and 1 were female. In the remaining other groups, there was male preponderance.

**Table V**  
**SEX DISTRIBUTION OF DIFFERENT TYPES OF CASES.**

Sex	TT	WT	WD	WL	WL	Total	Percentage
Male	8	9	11	9	16	59	83.07
Female	1	1	4	1	1	11	16.93
<b>Total</b>	<b>9</b>	<b>10</b>	<b>15</b>	<b>10</b>	<b>17</b>	<b>66</b>	<b>100.00</b>

### Clinical presentation of various types of cases :

Clinically the cases were divided in three groups namely tuberculoid, borderline and lepromatous. They had the following clinical features as shown in Table VI.

Table VI

#### CLINICAL PRESENTATION OF VARIOUS TYPES OF CASES.

Clinical features	Clinical diagnosis		
	Tuberculoid leprosy	Borderline	Lepromatous leprosy
No. of cases studied.	22	46	59
Raised margin and flat center with loss of sensation.	6	-	-
Raised thickened patches with loss of sensation.	4	-	-
Hypopigmented patches with partial loss of sensation.	22	6	-
Thickened nerve-sheath and/or peroneal.	14	7	10
Loss of sensation in peripheral part.	13	7	12
Deformities and trophic ulcers.	10	8	11
Loss of eye brows (Lateral 1/3)	8	5	?
Raised erythematous patches.	-	15	21
Macular lesions.	-	-	4

Hypomelanotic type : Out of 32 cases, 12 had hypopigmented patches of varying sizes all over body with partial loss of sensation for pain, temperature and touch. 6 cases showed flat lesions with thickened margins. They were receiving DDS for 3 to 5 years. 4 cases had thickened, raised, erythematous, anaesthetic patches, variable in size. Out of 32 cases, 14 had thickened nerve-fiber and/or peroneal, while 13 had loss of sensation in peripheral parts too. 10 cases were having deformities in the form of absorption of tip of fingers and toes with trophic ulcers. 8 were having loss of lateral 1/3 of eye brows.

Border line : 14 cases were studied in this group. 8 cases had raised, erythematous patches with shining surface, multiple in number and varying in size, present all over the body. 6 cases were also having hypopigmented patches with partial loss of sensation for pain, temperature and touch. 7 cases were having thickened ulnar and/or peroneal nerves with loss of sensation in peripheral parts. 8 cases were having deformities in the form of absorption of tips of fingers and toes. 8 cases were having loss of lateral one third of eye brows.

Irritative type : 29 cases were studied in this group. Out of which 21 were having erythematous bilaterally symmetrical patches of varying sizes with shining surface. 4 cases had given the history that they had erythematous

lesions 5 to 10 years back but had subsided after taking DDS for 5 to 10 years. These patients had extensive skin and/or perineal nerves. 6 cases were also having thickened nerves with other lesions. 15 cases were having loss of sensation in peripheral parts and 11 cases were associated with deformities and trophic ulcers. 7 cases were having loss of lateral one third of eye brows.

All cases were under treatment with DDS for 1-12 years duration. Skin biopsies were done and histopathological typing was done according to Ridley and Jopling (1962).

#### Clinical and histopathological correlation:

The clinical and histopathological correlation is shown in Table VII.

Table VII

#### CLINICAL AND HISTOPATHOLOGICAL CORRELATION.

Clinical Type	Histopathological Type						Total
	W	M	B	H	T	Total	
Tuberculoid Type.	9	11	2	1	1	33	
Borderline	-	3	9	3	-	18	
Lepromatous Type.	-	-	4	8	17	39	
Total	9	36	15	10	37	66	

Tubercloid type : Out of 22 cases, clinically classified as tubercloid type, 9 were tubercloid type (Fig. 2 a,b), 11 were borderline tubercloid (Fig. 2 a,b) and 2 were borderline on the basis of histopathological findings.

Borderline: Out of 16 cases, clinically classified as borderline, 3 were of BB, 9 were of BB (Fig. 4 a,b) and 3 were of BL type on the basis of histopathological findings.

Inflammatory type : Out of 39 cases clinically diagnosed as inflammatory type, 4 were of BB, 8 were of BL (Fig. 5 a,b) and 27 were of LL type (Fig. 6 a,b).

Variation in the presence of Myco. leprae in nasal smear, slit skin smear, and skin biopsy :

Variation in the presence of Myco. leprae in slit skin smear, nasal smear and skin biopsy is shown in Table VIII.

Table VIII  
VARIATION IN THE PRESENCE OF MYCO. LEPRAE IN NASAL SMEAR,  
SLIT SKIN SMEAR, AND SKIN BIOPSY.

Histo-pathological type.	Total No. of cases	Nasal smear examined	Positive rate for AFB	Slit skin smear examined	Positive rate for AFB	Biopsy performed	Positive rate
TT	9	9	-	9	-	9	-
BB	16	10	-	10	-	14	8
BB	15	8	1	8	-	15	7
BL	10	10	0	10	0	10	0
LL	27	9	0	9	0	27	0

*Mycobacterium tuberculosis* : All cases were negative for *Mycobacteria* in nasal smear, slit skin smear, and skin biopsies.

*Mycobacterium tuberculoides* : Out of 16 cases, in 10 cases the slit skin smears and nasal smears were observed and found to be negative, while in 6 skin biopsies, 5 were positive for *Mycobacteria*.

*Borderline* : Out of 16 cases, in 8 cases nasal and slit skin smears were prepared. Only 1 case of nasal smear was positive for *Mycobacteria* while all cases of skin slit smears were negative. Out of 15 skin biopsies, 7 biopsies were positive for AFB in (+).

*Borderline lepromatous* : Out of 10 cases, 3 cases were positive for *Mycobacteria* in nasal smear, 2 cases were positive in slit skin smear and 6 cases were positive in skin biopsies varying in grade from + to +++ (Fig. 7 a).

*Lepromatous leprosy* : 9 cases were examined for nasal and slit skin smear, out of which 4 cases and 3 cases were positive for AFB respectively. Out of 17 skin biopsies done, leprosy bacilli were present in 13 cases, varying in grade from + to +++++ (5+) (Fig. 7 b).

#### T-CELL COUNT IN VARIOUS TYPES OF LEPROSY AND CONTROL CASES :

Findings of absolute lymphocyte and T-cell count in various types of leprosy patients and

controls are given in Table IX (Fig. 8 and 9).

Table IX

DISTRIBUTION OF ABSOLUTE LYMPHOCYTE AND T-CELL COUNT IN DIFFERENT TYPES OF LEPROSY AND CONTROL CASES.

Type of cases	Absolute lymphocyte count, Mean $\pm$ SD ( Range )	T-cell % Mean $\pm$ SD ( Range )	Absolute T-cell count Mean $\pm$ SD ( Range )
Control	$2788.76 \pm 696.48$ (1723 - 4408)	$66.88 \pm 5.11$ (58 - 77)	$1856.86 \pm 810.47$ (2973 - 3226)
Tuberculosis Type	$3089.77 \pm 747.74$ (2207 - 5043)	$61.66 \pm 2.85$ (58 - 67)	$1993.09 \pm 830.40$ (1497 - 3297)
Borderline tuberculosis	$2695.71 \pm 693.96$ (2490 - 3200)	$65.89 \pm 7.01$ (46.5 - 78)	$2095.93 \pm 766.46$ (1190 - 6100)
Borderline	$3777.60 \pm 615.99$ (2276 - 5222)	$68.86 \pm 4.77$ (60 - 67)	$1668.53 \pm 361.96$ (1160 - 2778)
Borderline Lepromatous	$2274.4 \pm 806.03$ (1040 - 3480)	$48.85 \pm 5.37$ (41 - 59)	$1609.8 \pm 870.71$ (631 - 2671)
Lepromatous Type	$2898.82 \pm 985.64$ (1575 - 4250)	$68.39 \pm 6.02$ (41 - 87)	$1578.55 \pm 891.46$ (610 - 3465)

In control cases, absolute lymphocyte counts ranged from 1723 to 4408 with mean ( $\pm$  SD) of 2788.76 ( $\pm$  696.48). T-cell percentage ranged from 58 to 77 with mean ( $\pm$  SD) of 66.88 ( $\pm$  5.11). Absolute T-cell count ranged from 2973 to 3226 with mean ( $\pm$  SD) of 1856.86 ( $\pm$  810.47).

In tuberculous type of cases absolute lymphocyte count ranged from 3037 to 3015 with mean ( $\pm SD$ ) of 3009.77 ( $\pm 747.74$ ). T-cell percentage ranged from 66 to 67 with mean ( $\pm SD$ ) of 61.66 ( $\pm 2.03$ ). Absolute T-cell count ranged from 1497 to 3307 with mean ( $\pm SD$ ) of 1992 ( $\pm 630.49$ ).

In borderline tuberculous type of cases absolute lymphocyte count ranged from 3130 to 3536 with mean ( $\pm SD$ ) of 3695 ( $\pm 893.96$ ). T-cell percentage ranged from 46.5 to 78 with mean ( $\pm SD$ ) of 55.09 ( $\pm 7.01$ ). Absolute T-cell count ranged from 1192 to 4188 with mean ( $\pm SD$ ) of 2898.93.

In borderline type of cases, absolute lymphocyte count ranged from 2876 to 3002 with mean ( $\pm SD$ ) of 2777.60 ( $\pm 618.99$ ). T-cells percentage ranged from 50 to 67 with mean ( $\pm SD$ ) of 55.36 ( $\pm 4.77$ ). Absolute T-cell count ranged from 1160 to 2778 with mean ( $\pm SD$ ) of 1665.33 ( $\pm 504.96$ ).

In borderline lepromatous type of cases, absolute lymphocyte count ranged from 1940 to 2400 with mean ( $\pm SD$ ) of 2274.4 ( $\pm 624.04$ ). T-cell percentage ranged from 41 to 59 with mean ( $\pm SD$ ) of 49.35 ( $\pm 5.37$ ). Absolute T-cell count ranged from 831 to 1871 with mean ( $\pm SD$ ) of 1409.8 ( $\pm 270.71$ ).

In lepromatous type of cases, absolute lymphocyte count ranged from 1775 to 4050 with mean ( $\pm SD$ )

of 2288.00 ( $\pm 988.64$ ). T-cell percentage ranged from 41 to 57 with mean ( $\pm SD$ ) of 46.00 ( $\pm 4.00$ ). Absolute T-cell count ranged from 636 to 3496 with (mean  $\pm SD$ ) of 1370.00 ( $\pm 591.46$ ).

**Significance T-cell percent between different groups:**

Significance of the difference of T-cell percent between different groups is shown in Table X.

The statistical significance in the difference of T-cell percentage is calculated between control and total leprosy cases and has been observed to be highly significant (P-value  $<.001$ ). The significance in the difference of T-cell percentage is also calculated between control and different type of leprosy cases separately and has been observed to be highly significant (P-value  $<.001$ ) in all types. T-cell percentage is gradually decreased from TT to LL. The differences in T-cell percentage between TT and BT, TT and BB, TT and BL, TT and LL, is calculated and has been observed to be statistically highly significant (P-values  $<.001$ ) except between TT and BT, where it is only significant (P-value  $<.05$ ).

Table X

SIGNIFICANCE OF T-SUM PERCENT BETWEEN DIFFERENT GROUPS

T-SUM PERCENT (%)	NUMBER OF SUBJ. (n)	T-SUM % (MEAN ± SD)	Statistical significance		P-value
			Difference between groups	Difference between n groups	
0.00	22	65.88 ± 5.14 (65.88 ± 7.77)	-	-	-
1.00	22	71.55 ± 7.33 (71.55 ± 6.67)	central and impurity areas	$L_{.001}$	-
2.00	22	61.66 ± 3.99 (61.66 ± 6.67)	c and $\pi$	$L_{.001}$	-
3.00	22	67.09 ± 7.09 (67.09 ± 7.09)	c and $\pi$	$L_{.001}$	$\pi\pi$ and $\eta\eta$
4.00	22	67.36 ± 5.77 (67.36 ± 6.67)	c and $\pi$	$L_{.001}$	$\pi\pi$ and $\eta\eta$
5.00	22	69.48 ± 5.37 (69.48 ± 6.67)	c and $\pi$	$L_{.001}$	$\pi\pi$ and $\eta\eta$
6.00	22	70.88 ± 5.09 (70.88 ± 6.67)	c and $\pi$	$L_{.001}$	$\pi\pi$ and $\eta\eta$
7.00	22	71.20 ± 5.37 (71.20 ± 6.67)	c and $\pi$	$L_{.001}$	$\pi\pi$ and $\eta\eta$
8.00	22	71.20 ± 5.37 (71.20 ± 6.67)	c and $\pi$	$L_{.001}$	$\pi\pi$ and $\eta\eta$
9.00	22	71.20 ± 5.37 (71.20 ± 6.67)	c and $\pi$	$L_{.001}$	$\pi\pi$ and $\eta\eta$
10.00	22	71.20 ± 5.37 (71.20 ± 6.67)	c and $\pi$	$L_{.001}$	$\pi\pi$ and $\eta\eta$

Significant differences  
 1.  $\pi\pi$  - central area  
 2.  $\pi\pi$  -  $L_{.001}$  significant  
 3.  $L_{.001}$  - impurity areas

B-CELL COUNT IN VARIOUS TYPES OF LEPROSY AND CONTROL CASES.

Findings of B-cell percentage and absolute B-cell count in various types of leprosy and control cases are given in Table XI (Fig.10 and 11).

Table XI

DISTRIBUTION OF B-CELL COUNT IN DIFFERENT TYPES OF LEPROSY AND CONTROL CASES.

Type of cases	B-cell percentage Mean $\pm$ SD (Range)	Absolute B-cell count Mean $\pm$ SD (Range)
Control	23.06 $\pm$ 3.62 ( 16 - 30 )	634.92 $\pm$ 50.81 ( 509 - 1101 )
Tuberculoid type	27.05 $\pm$ 3.16 ( 21 - 33 )	889.77 $\pm$ 297.08 ( 700 - 2680 )
Borderline tuberculoid	22.78 $\pm$ 7.09 ( 10 - 45 )	1191.64 $\pm$ 504.81 ( 660 - 1630 )
Borderline	24.76 $\pm$ 4.38 ( 18 - 34 )	1086.59 $\pm$ 309.91 ( 641 - 1976 )
Borderline lepromatous	30.2 $\pm$ 4.68 ( 23 - 45 )	1195.0 $\pm$ 212.86 ( 776 - 2636 )
Lepromatous type	43.5 $\pm$ 5.0 ( 33- 55.5 )	1245.64 $\pm$ 232.77 ( 660 - 1700 )

In control cases B-cell percentage ranged from 16-30 with mean ( $\pm$ SD) of 23.06 ( $\pm$ 3.62). Absolute B-cell count ranged from 509 to 1101 with mean( $\pm$ SD) of 634.92 ( $\pm$ 50.81).

In tuberculoïd type of cases, B-cell percentage ranged from 21 to 33 with mean ( $\pm SD$ ) of 27.08 ( $\pm 3.18$ ). Absolute B-cell count ranged from 753 to 1689 with mean ( $\pm SD$ ) of 1090.77 ( $\pm 397.03$ ).

In borderline tuberculoïd type of cases, B-cell percentage ranged from 30 to 45 with mean ( $\pm SD$ ) of 32.78 ( $\pm 7.09$ ). Absolute B-cell count ranged from 650 to 1630 with mean ( $\pm SD$ ) of 1101.64 ( $\pm 304.51$ ).

In border line type of cases, B-cell percentage ranged from 30 to 46 with mean ( $\pm SD$ ) of 34.76 ( $\pm 4.38$ ). Absolute B-cell count ranged from 641 to 1974 with mean ( $\pm SD$ ) of 1006.00 ( $\pm 300.91$ ).

In borderline lepromatous type of cases, B-cell percentage ranged from 30 to 45 with mean ( $\pm SD$ ) of 38.2 ( $\pm 4.62$ ). Absolute B-cell count ranged from 776 to 2496 with mean ( $\pm SD$ ) of 1285.8 ( $\pm 222.86$ ).

In lepromatous type of cases, B-cell count ranged from 35 to 55.5 with mean ( $\pm SD$ ) of 42.5 ( $\pm 5.0$ ). Absolute B-cell count ranged from 650 to 1700 with mean ( $\pm SD$ ) of 1145.64 ( $\pm 242.77$ ).

**Significane of B-cell percent in different group :**

Significane of the difference of B-cell count between control and different type of cells is shown in Table XIII.

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Type of series	No.	Mean $\pm$ standard error (in mm)	Statistical significance		
			Differences between various groups	p value	Difference between groups
control	35	37.05 $\pm$ 0.68 ( $n = 30$ )	-	-	-
total leucocyte counts	68	35.8 $\pm$ 0.38 ( $n = 55$ )	Control and total leucocyte counts.	$L < 0.001$	$L < 0.001$
WBC	9	37.38 $\pm$ 0.48 ( $n = 32$ )	C and R	$L < 0.001$	$L < 0.001$
	10	37.75 $\pm$ 0.69 ( $n = 35$ )	C and R	$L < 0.001$	$L < 0.001$
	11	37.76 $\pm$ 0.79 ( $n = 36$ )	C and R	$L < 0.001$	$L < 0.001$
	12	37.42 $\pm$ 0.68 ( $n = 35$ )	C and R	$L < 0.001$	$L < 0.001$
	13	37.45 $\pm$ 0.68 ( $n = 35$ )	C and R	$L < 0.001$	$L < 0.001$

There is marked difference in mean of B-cell percentage of the control group and total leprosy cases group. The greatest increase in mean percentage of B-cell is observed from TT to LL. The statistical significance is calculated between control and total leprosy cases, control and TT, control and BB, control and BL, and control and LL, has been observed to be highly significant ( $P$ -value  $< .005$ ) statistically in all groups. The difference in B-cell percentage is also calculated between TT and BB, TT and BL, TT and LL and LL and BL and has been observed to be highly significant ( $P$ -value  $< .005$ ) statistically except TT and BB, where it is significant only ( $P < .05$ ).

#### LYMPHOBLAST TRANSFORMATION PERCENTAGE IN VARIOUS TYPES OF LEPROSY AND CONTROL CASES :

Findings of lymphoblast transformation response to PHA in various types of leprosy and control cases is shown in Table XXX (Fig. 12 and 13).

Table XIII

DISTRIBUTION OF LYMPHOBLAST TRANSFORMATION PERCENTAGE  
IN DIFFERENT TYPES OF LEPROSY AND CONTROL GASES.

Type of cases	Lymphoblast transformation percentages. Mean $\pm$ SD ( Range )
Control	38.43 $\pm$ 3.05 ( 34 - 46.8 )
Tuberculoïd type	36.45 $\pm$ 3.46 ( 32 - 40.8 )
Borderline tuberculoïd	37.07 $\pm$ 4.00 ( 31.5 - 50 )
Borderline	38.37 $\pm$ 3.69 ( 30 - 39.8 )
Borderline lepromatous	30.06 $\pm$ 3.81 ( 14.5 to 30.5 )
Lepromatous	22.47 $\pm$ 2.89 ( 9 - 25.8 )

Lymphoblast transformation percentage in control cases ranged from 36 to 46.5 with mean ( $\pm SD$ ) of 38.12 ( $\pm 3.66$ ). Lymphoblast transformation percentages in TT, MT, BB, DL and LL ranged from 36 to 40.5, 31.5 to 38, 30 to 30.5, 14.5 to 20.5 and 9 to 15.5 and their mean ( $\pm SD$ ) are 38.12 ( $\pm 3.66$ ), 34.48 ( $\pm 5.44$ ), 27.57 ( $\pm 4.29$ ), 23.57 ( $\pm 3.69$ ), 18.08 ( $\pm 3.31$ ) and 12.47  $\pm$  2.39 respectively.

**Significance of lymphoblast transformation percentage in various groups :**

Significance of difference of lymphoblast transformation percentage is shown in Table XIV.

Lymphoblast transformation percentage is markedly reduced between control group and total leprosy group. The gradual decrease in percentage of lymphocyte blast transformation is observed from TT to LL. The differences in control and total leprosy group, and control and different type of leprosy groups are calculated and has been observed to be highly significant ( $P < .005$ ) except control and TT, where it is significant only ( $P < .05$ ). The differences between TT and other types is also calculated separately and has been observed to be highly significant in all types of cases ( $P < .005$ ).

Table XXV

## SIGNIFICANCES OF TRANSPORTATION PERCENT BETWEEN DIFFERENT GROUPS

Groups of series (c)	No. of cases studied	Mean $\pm$ S.E. (Range)	Statistical significances		
			Differences between groups	t values	P values between groups
control	32	28.13 $\pm$ 3.06 (35 - 46.5)	-	-	-
total Impover- ished cases	62	28.36 $\pm$ 0.30 (29 - 40.0)	Control and total Impover- ished cases.	-	$< .001$
7	28.45 $\pm$ 3.41 (25 - 40.5)	c and w	-	-	$< .05$
11	27.57 $\pm$ 4.00 (24.5 - 35)	c and w	-	-	$< .001$
15	28.57 $\pm$ 3.59 (26 - 36.5)	c and w	-	-	$< .001$
10	28.48 $\pm$ 3.21 (26.5 - 30.5)	c and w	-	-	$< .001$
17	28.47 $\pm$ 3.29 (29 - 35.5)	c and w	-	-	$< .001$
			w = statistical significance	$> .7$	statistical significance
				$< .01$	$< .01$ statistical significance

Thus it is observed that there is gradual decrease in T-cell percentage and lymphoblast transformation percentage from TT to LL. On the other hand, gradual increase in B-cell percentage is observed from TT to LL.

**Response of lepromin and candida antigens in control and study group :**

The response of lepromin and candida antigens is shown in Table XV.

**Table XV**  
**RESPONSE OF LEPROMIN AND CANDIDA ANTIGENS IN CONTROL AND DIFFERENT TYPES OF LEPROSY CASES.**

Type of cases.	No. of cases examined.	Lepromin test		Candida test	
		No. of cases positive	Percentage	No. of cases positive	Percentage
Control	50	32	66.6	7	35.0
Tuberculoïd type.	9	6	66.6	1	11.1
Borderline tuberculoïd	24	6	43.8	3	21.4
Borderline	15	6	40	3	20.0
Borderline lepromatous	10	-	-	-	-
Lepromatous leprosy.	17	-	-	-	-

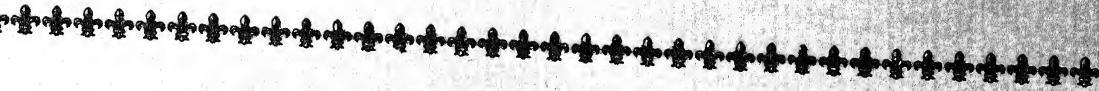
**Central Leprosy :** Out of 28 cases, 18 were inoculated with Phasmadis leprosin and candida antigens. 12 cases (66.6%) were positive for early (Pernicious) reaction and 7 (38.8%) were positive for candida antigen.

**Tuberculoid :** 9 cases were inoculated with Phasmadis leprosin and candida antigen. 6 cases (66.6%) were positive for early (Pernicious) reaction and 4 cases (44.4%) were positive for candida antigen. One case turned up on 21st day showing ulceration at site of inoculation.

**Borderline tuberculoid :** 16 cases were inoculated with Phasmadis leprosin and candida antigens. 6 cases (43.8%) were positive for early (Pernicious) reaction and 3 cases (21.4%) were positive for candida antigen. One case turned up on the 21st day showing nodule formation.

**Borderline :** 15 cases were inoculated with Phasmadis leprosin and candida antigens. 6 cases (40%) were positive for early (Pernicious) reaction and 3 cases (20%) were positive for candida antigen.

**Borderline lepromatous and lepromatos type:** 10 cases of borderline lepromatoses and 17 cases of lepromatos type were inoculated for Phasmadis leprosin and candida antigen. None were positive for either antigen.



## DISCUSSION



It is a fact that although leprosy is an infectious disease, a large majority of population goes through a stage of sub-clinical infection. Most of these people are able to mount resistance towards *Mycobacterium leprae* infection. A minor group of population, due to unknown mechanism, is unable to mount a proper immune response against *Mycobacterium leprae* invasion.

The present study was carried out to study cell mediated immunity in different types of leprosy and control cases. The total number of cases were 90 which included 63 cases of different type of leprosy and 27 cases of control. The leprosy cases were classified histopathologically into tuberculoid type-9, borderline tuberculoid type-1A, borderline type-1B, borderline lepromatous type-1D and lepromatous type-1E.

T-cell count, B-cell count, lymphoblast transformation percentage, skin tests using leprosin and candida antigen; and nasal, slit-skin smear, and skin biopsies for acid fast bacilli were studied. Uniformity of the study has been maintained by studying the leprosy cases and age and sex matched control cases. The majority of cases were within the third to fifth decades with mean age ( $\pm SD$ ) of 30.27 ( $\pm 10.68$ ). The control cases were almost of similar age group with mean ( $\pm SD$ ) of 30.5 ( $\pm 9.99$ ).

In lepromatous leprosy male, female ratio is generally reported as being around 2:1 and in tuberculoid no sex differentiation has been found (Anderson and Kissane, 1977). Similar incidence was also observed in present study as 5 were male out of 9 cases of tuberculoid type while in other types male preponderance was noted. Variation in the presence of Myco. leprae in nasal smear, slit-skin smear and skin biopsy :

Myco. leprae has special affinity for skin, nerve tissue and nasal mucosa. It may be demonstrated in skin-slit smear, nasal smear and skin biopsy. In present study, 2 cases were positive for Myco. leprae in skin-slit smears out of 10 and 17; BL and LL cases respectively while positive nasal smears in BB, BL and LL were 1, 3 and 4 out of 18, 10 and 9 respectively. The skin biopsies were positive in all types of leprosy except tuberculoid. The number of positive cases in skin biopsy for Myco. leprae in BT, BB, BL and LL were 5, 7, 6 and 10, out of 16, 15, 10 and 13 cases respectively.

It is evident from above findings that the number of positive cases were less in nasal and slit-skin smear as compared to skin biopsy. This may be explained as patient were under treatment with DDS for longer duration and bacilli may have cleared from skin and invaded the internal organs.

## T-CELL VARIATION IN CONTROL AND DIFFERENT TYPES OF LIVERD CASES :

### Control Group :

In the present study, 26 age and sex matched control cases have been studied. T-cell percentage ranged from 53 to 77 with mean ( $\pm$  SD) of 66.08 ( $\pm$  5.15). Absolute T-cell count ranged from 1229 to 2979 with mean ( $\pm$  SD) of 2066 ( $\pm$  310.47).

Similar were the observations of Rao et al (1976) who had reported mean ( $\pm$  SD) of T-cell percentage 68.8 ( $\pm$  7.7). Other workers also have reported nearly similar mean T-cell percentage which ranged from 40.8 to 77 percent (Dwyer et al, 1978; Liu et al, 1974; Chogla et al, 1977; Sharma et al, 1979).

There are various factors which affects T-cell percentage in normal human peripheral blood. Mendes et al (1973) had studied the conditions in E and EAC rosette formation in great detail for T and B-cells respectively. They had found that E and EAC rosettes obtained at Duke University were 55.4 percent and 18.3 percent and at Escola Paulista de Medicina were 30.4 and 26.2 percent respectively. Rosette formation between T-cells and sheep red blood cells was found to be temperature dependent with maximum values between 10°C to 25°C and no rosette formation at 37°C. They had also mentioned that variation in rosette percentage was also observed

when same normal donor was tested on different days and this is also affected by storage of blood and/or lymphocyte.

Therefore to minimize these effects, in present study the fresh samples were used and tests were carried out at 4 to 8°C. From the above mentioned facts, it is clear every laboratory should have its own control value for comparison with study cases. The minor differences in the present study and other studies may be due to above mentioned facts.

#### Lymphocyte Count :

The absolute lymphocyte count was almost equal in all types of leprosy. The mean ( $\pm SD$ ) of absolute lymphocyte count in TT, MT, BB, DL and LL was 3069.77 ( $\pm 747.47$ ), 3695.71 ( $\pm 899.96$ ), 2777.60 ( $\pm 615.99$ ), 2274.4 ( $\pm 304.06$ ) and 2220.00 ( $\pm 955.64$ ) respectively. It indicates that there is no correlation in absolute lymphocyte count among different groups.

The mean T-cell percentage ( $\pm SD$ ) in control was 66.00 ( $\pm 5.11$ ) which was decreased to 45.50 ( $\pm 4.08$ ) in LL as shown in Table IX. Similarly there was decrease in absolute T-cell count towards lepromatous pole. The mean T-cell percent was decreased in all types of leprosy cases. The fall in mean T-cell percentage was maximum in LL and minimum in TT as compared to control cases. In case of intermediate pole while MT was closer to TT and DL was closer to LL.

The difference in mean T-cell percentage between control and total leprosy cases was statistically highly significant ( $P = <.005$ ) and was significant also when control cases were compared with different type of leprosy cases separately. The difference in TT and BB was only significant ( $P <.05$ ) but it was highly significant with BB, BL and LL ( $P = <.005$ ).

Other workers had also reported the similar findings that there were decrease in mean T-cell percent from control to LL (Soyer et al, 1973; Lin et al, 1974; Chagle et al, 1977; Kallmann et al, 1977).

Recently Sharma et al (1979) had studied a complete spectrum of leprosy. They had combined the TT and BB group and found that mean ( $\pm SD$ ) of T-cell percentage in control, TT + BB, BB, BL and LL cases which were 55.87 ( $\pm 7.38$ ), 51.37 ( $\pm 10.45$ ), 40.76( $\pm 8.48$ ), 39.88 ( $\pm 7.3$ ) and 37.00 ( $\pm 11$ ). It indicates that there was gradual decrease in the mean T-cell percent from control to LL which is similar to present study with minor differences which may be due to factors mentioned earlier.

Contrary to the findings of present study, Rao et al (1976) had observed that there was no difference in mean T-cell percent in control and LL. They had found mean ( $\pm SD$ ) of T-cell percent which were 70.4 ( $\pm 6.3$ ) and 68.8 ( $\pm 7.7$ ) respectively.

The non specific depletion of cell mediated immunity can be due to an absolute reduction of T-cells. The specific defect of lymphocyte in leprosyous patients producing tolerance to Myco. leprae continued to be present even after all demonstrable bacilli are eliminated.

Dupper et al (1970) has suggested two mechanism for alteration in T, B ratio. First was that the para-cortical, area which was normally heavily populated with T-cell, were often extensively invaded by macrophage laden bacteria. Secondly presence of antigen within granulomas of these disease for a longer period of may induce sustained production of tissue-suppressive factors resulting in anergy.

Kohlmann et al (1977) had suggested that in bacteriologically negative DL cases, the mean value for circulating lymphocyte were not significantly different from those of normal control. Thus T-lymphocyte deficiency did not appear to be genetic because number of B-lymphocytes returned to normal in bacteriologically negative DL patients. T-cell deficiency which may be the reason for diminished cell mediated immune response, is further supported by prolonged survival of skin allografts in leprosy patients (Das et al, 1971).

#### B-cell variation in control and leprosy cases:

B-cell percentage was gradually increased from control to DL. The mean ( $\pm$  SD) of B-cell percentage in control was 28.86 ( $\pm$  8.68) and gradually increased up to 39.5 ( $\pm$  8.9) in leprosyous leprosy as shown in Table XII.

The difference in B-cell percentage in control and total leprosy cases as well as different type of leprosy cases separately is shown in Table XIII. The difference was highly significant in control and total leprosy cases ( $P = \underline{<.005}$ ). It was only significant ( $P = \underline{<.05}$ ) in control and TT but was highly significant ( $P = \underline{<.005}$ ) in other types. When TT was compared with BT, BB, BL and LL, it was highly significant in all except with BT where it was only significant ( $P = \underline{<.02}$ ).

Garg and Poonawala et al (1973) had reported marked increase in B-cell percentage 60% to 65% of B-cell in peripheral blood. Sharma et al (1979) had studied B-cell percentage in complete spectrum of leprosy and observed slight increase in B-cell percentage. They observed mean ( $\pm SD$ ) of B-cell percentage in control, TT + BT, BB, BL and LL which was 27.67 ( $\pm 3.77$ ), 28.50 ( $\pm 5.71$ ), 30.23 ( $\pm 7.09$ ), 31.06 ( $\pm 7.01$ ) and 29.97 ( $\pm 7.97$ ) and concluded that there was very minimal increase in B-cell percentage which was statistically insignificant. Rao et al (1976) had also observed no significant difference in increase of B-cell in lepromatous leprosy.

Deyar et al (1973) had studied B-cell percentage only in control and lepromatous groups and found significant increase in B-cell percentage in peripheral blood in lepromatous group. They observed 27 percent in control and 38 percent B-cell in lepromatous group similar to the

observation of present study. Other workers also had shown that patients with lepromatous leprosy had high proportion of circulating lymphocytes possessing membrane bound immunoglobulin ( B-cell ). It had been proposed that such increase in B-cell numbers might represent over compensation for a deficiency of T-lymphocytes.

The findings of present study were also in accordance with Chogle et al (1977), who studied Mean ( $\pm$ SD) of B-cell percentage in control, TT, BB and LL groups and found 27 ( $\pm$ 5.4), 36 ( $\pm$ 8.6), 37 ( $\pm$ 6.6) and 36 ( $\pm$ 10.7) respectively.

The findings of present study were again supported by the study of Wane et al (1971) who had observed significant increase in B-cell percentage, lymphocytes obtained from crushing the lymphnode.

As B-lymphocytes are involved in antibody production. Abe et al (1972) had reported anti Myco. leprae antibodies with indirect fluorescent technique in both lepromatous, tuberculoid, and indeterminate cases but the proportion of positive sera and titres observed was highest in lepromatous sera.

It had established that human antibodies produced by B-cell could specifically enhance or inhibit the response of T-cells (cited by Chogle et al, 1977).

It had also been suggested that T-cell might be affected by depressive humor factor (Butlock, 1968).

#### LYMPHOBLAST TRANSFORMATION RESPONSE TO PMA IN CONTROL, CASES AND LATENT CASES.

Blood lymphocytes were found to be stimulated to blastogenesis by mitogen such as PMA and could be a measure of cell mediated immune response ( David and David, 1972). In present study, there was gradual decrease in lymphoblast transformation to PMA from control to LL. The mean ( $\pm$  SD) of lymphoblast transformation percentage in control, TT, BB, DL and LL was 36.12 ( $\pm$  3.06), 34.45 ( $\pm$  3.44), 37.87 ( $\pm$  4.38), 23.37 ( $\pm$  3.69), 26.05 ( $\pm$  3.21) and 23.47 ( $\pm$  3.29) respectively as shown in Table XIII.

The difference in lymphoblast transformation response to PMA in control and total leprosy cases as a whole and separately with different type of leprosy cases were found to be highly significant ( $P = <.001$ ) except control and TT where it was only significant ( $P = <.05$ ). The difference between TT and DL, TT and BB, TT and DL and TT and LL was also highly significant ( $P = <.005$ ). It indicates that there was marked decrease in lymphoblast transformation response towards lepromatous pole.

The findings of the present study were almost similar to the usually reported findings of Choi et al (1980) and Dubey et al (1981) who had also observed the diminished lymphogenic response to PMA with slight variation. Choi et al (1980) had reported mean ( $\pm$  SD) of lymphoblast transformation percentage in control, TT, BT, BB, BL and LL which were 34.6 ( $\pm$  8.8), 20.6 ( $\pm$  10.2), 20.9 ( $\pm$  6.8), 19.1 ( $\pm$  12.8) and 16.7 ( $\pm$  10.8) respectively. Dubey et al (1981) had reported the percentage of lymphoblast transformation which had decreased from 33 to 40 percent in TT to 8 to 30 percent in LL.

These findings were further supported by the findings of other workers who had also studied the lymphoblast transformation response to various other antigens by measuring the DNA synthesis of cells by radioactive thymidine uptake. They had shown diminished response from TT to LL (Kao et al, 1971; Furt Chingwong et al, 1971; Mohra et al, 1973; Tahar et al, 1973; Durr et al, 1973; Job et al, 1976; Kallmann et al, 1977; and Shams et al, 1979).

However Ulrich et al (1978) found no significant difference in control, tuberculous, leprosy-like, and lepromatous patients to their responses to PMA and Fuks wood antigen (FWA).

Although these findings were much closer to the findings of other worker with slight variation, this discrepancy in the findings of PHA stimulation may be due to the effects of drugs received during the study. Ganguly et al (1979) had observed diminished response in healthy volunteers after receiving the DDS orally. Similar were the observations of Beijnen and Pienai (1974) after studying the effect of DDS in vitro.

It has been suggested that T-cell number is significantly reduced towards lepromatous pole as shown in present as well as in other studies too. Hence, reduced lymphoblast transformation response to PHA may be due to less number of T-cells towards lepromatous end of spectrum. TT and LR patients do not show such reduction in T-cells and thus probably showed slight diminution in lymphoblast transformation response to PHA.

Further Bullock and Pienai (1971) had observed the presence of depressor activity to blastogenesis in plasma of leprosy cases who also claimed that depressor activity did not get altered by antileprosy treatment. In contrast to this Dabey et al (1961) had shown that there was significant improvement in their blast transformation percentage in LL patients taking DDS over untreated LL cases.

Nelson et al (1971) had reported that the lepromatous patient as a whole had no evidence of intrinsic defect in the ability of lymphocytes to respond to PHA. Lymphocyte from Indian lepromatous patients with stable disease did show a depressed response when cultured in normal reference serum. In unselected Malay and Indian lepromatous patients a depressed response was clearly apparent only when the cells cultured in autologous serum. In the Chinese lepromatous patients the lymphocyte actually responded better than those of normal Chinese even in autologous serum. Thus there was racial difference in response to PHA induced transformation.

#### Response of Ipponiois and Candida antigen in Control and Leprosy Group :

The findings of the present study showed that leprosy test was positive in 77, 37 and 33 while RL and LL patients failed to respond to leprosy antigen. Similar results were obtained with candida antigen with the difference that percentage of positive cases were less. The percentage of positive leprosy cases in control, 77, RL, LL were 66.6, 66.6, 43.8 and 40 respectively while with candida were 28.6, 44.4, 21.4 and 20 respectively.

Bach et al (1968) had only used candida antigen for testing delayed hypersensitivity reaction in their study of control, tuberculous and lepromatous cases. They had also observed diminished response in leprosy cases both in tuberculous as well as in lepromatous type and reported that positive response against candida were seen in 40.7%, 10.6% and 14.5% in control, tuberculous and lepromatous type respectively. The discrepancy in present study and study of Bach et al (1968) who had observed positive cases even in lepromatous type, may be due to the fact that they had divided the cases only in two groups namely tuberculous and lepromatous group. So some of borderline type of cases may be included in the lepromatous type who had given the positive response.

From the findings of present study, it is clear that lepromin test is positive upto 33 cases, and this may be a good measure of immunity during the course of therapy or during the reversal reactions, when patient is shifting from one pole to another pole.

Thus it is established from the present study that there is gradual decrease in T-cell percentage, PHA response, and cutaneous delayed hypersensitivity tests from tuberculous pole to lepromatous pole. Borderline group comes in between the two polar forms in the Ridley Jeppling scale. It comes closer to

TT and ML cases closer to L. D-sell percentage is gradually increased from tubercloid to lepromatous pole. The biopsies are more important for demonstration of Myco-lepros than slit-skin smear and nasal smear.

Bullock et al (1968) had studied the leprosia response in lepromatous and tubercloid cases and found 20 positive cases out of 56 and 28 positive out of 53 cases respectively. This controversy may be due to the fact that they had divided the cases in two groups only. So, it may be possible that few borderline cases may have been included in lepromatous group giving percentage of positive response. They had also studied the response of canthide in lepromatous and tubercloid type and found 24 positive cases out of 56 and 27 out of 53 respectively. The same reason may be also in this. They had also studied hypersensitivity response with various other antigens such as PPD, trichophytes and chemical like picryl chloride, and found diminished response in lepromatous patients than of tubercloid. They had also compared treated and untreated cases and found better response in treated cases as compared to untreated. Kumar (1980) had observed similar results.



## CONCLUSIONS



The present study was conducted on 90 cases including 35 control and 65 different types of leprosy patients from June 1981 to March 1982. Out of 65 leprosy patients, 47 were D; 17 were M; 22 were BB; BL were 10 and LL were 17 on the basis of histopathological criteria of Ridley and Jopling.

The majority of cases (48) were in third to fifth decade with male preponderance except in tuberculoid type where male-female ratio was almost equal.

T-cell & B-cell & lymphoblast transformation response to PMA, skin tests using lepromin and candid antigen were done in all cases. T-cell were studied by rosette formation with un sensitized sheep red blood cells; B-cells were studied by rosette formation with antibody and complement coated sheep red blood cells. Lymphoblast transformation response to PMA was observed after 3 days culture in TC medium 199 after staining by Leishman's stain.

These studies had shown that there was diminished cell mediated immune response in various types of leprosy being maximum in lepromatous and minimum in tuberculoid type in Ridley-Jopling scale.

It was observed that mean ( $\pm$ SD) of T-cell percentage in control, TT, MT, BB, BL and LL were 66.33 ( $\pm$ 5.41), 61.66 ( $\pm$ 3.89), 55.09 ( $\pm$ 7.01), 55.36 ( $\pm$ 4.77), 48.38 ( $\pm$ 5.37) and 45.38 ( $\pm$ 4.08) respectively.

On contrary, B-cell count was observed to increase gradually from tuberculoid pole to lepromatous pole. The mean ( $\pm SD$ ) of B-cell percentage in control, TT, BT, BB and LL were 22.05 ( $\pm 3.62$ ), 27.05 ( $\pm 3.18$ ), 32.75 ( $\pm 7.09$ ), 34.75 ( $\pm 4.38$ ), 38.2 ( $\pm 4.68$ ) and 43.8 ( $\pm 5.0$ ) respectively.

The lymphoblast response to PHA was found to decline gradually from tuberculoid to lepromatous-pole like decrease in T-cell percentage. The mean ( $\pm SD$ ) of percentage of lymphoblast transformation response in control, TT, BT, BB and LL were 38.18 ( $\pm 3.05$ ), 34.45 ( $\pm 5.44$ ), 27.57 ( $\pm 4.33$ ), 23.37 ( $\pm 3.69$ ), 19.05 ( $\pm 3.31$ ) and 12.47 ( $\pm 3.39$ ) respectively.

The skin tests using candida and leprosin were also performed in control as well as leprosy cases. The percentage of positive cases with leprosin antigen in control, TT, BT, and BB were 66.6, 66.6, 42.8 and 40 and with candida antigen were 38.2, 44.4, 31.4 and 20.0 respectively. BL patients did not respond to such extent that they could be labelled as positive while LL patient failed to respond with either antigen.

The following conclusions were drawn from the present study :

- (1) The incidence was high in male in all type of leprosy except in tuberculoid type where it was almost equal in both sexes.

- (a) There was fall in percentage of T-cell in all types of leprosy cases. The fall being maximum in LL and minimum in TT. The borderline group showed T-cell percentage in between the polar forms.
- (b) Absolute T-cell count was diminished in BB, BL and LL while almost equal in TT and BT with control.
- (c) There was gradual increase in B-cell percentage from tuberculoid pole to lepromatous pole. The increase in B-cell percentage was more marked in LL and least marked in TT. BB was in between the polar forms.
- (d) Lymphoblast transformation response to PHA was decreased from TT to LL similar to T-cell percentage.
- (e) Cutaneous response to leprosin and candida antigen were positive in TT, BT and BB while negative in BL and LL. The percentage of positive cases with leprosin was more as compared to candida antigen.

From the present study, it is concluded that there is gradual fall in cell mediated immune response from tuberculoid pole to lepromatous pole. Borderline group lies at mid position of spectrum while BT is closer to TT and BL is closer to LL. The above mentioned tests are useful to assess the cell mediated immune status of patients which are very important during reversal reactions and during course of therapy when patients may shift on either side of spectrum in BT, BB, and BL.

## BIBLIOGRAPHY

1. Abo,M., Minagawa,F., Yoshida,Y. and Okamura,K. : Studies on antigenic specificity on *Mycobacterium leprae*. II Purification and immunological characterization of soluble antigen in leprosy nodules. Int. J. Lepr., 49:107, 1978.
2. Anderson,J.G. : Studies in the medieval diagnosis of leprosy, a thesis for the Doctorate in Medicine, published as supplement to Davis medical bulletin, Vol. 16, 1969 ( Quoted by Bhargava & Leprosy Ed.I Kothari Med. Pub. House, Bombay, pp 12, 1978).
3. Anderson,W.A.B. and Kissane,J.M. : Pathology. Vol. I, Ed. 17th. Pub. C.V. Mosby Comp St Louis Missouri, pp 418, 1977.
4. Arnold,N.L. : Polar concept in leprosy. Int. J. Lepr. 42 : 489, 1974.
5. Barbieri,T.A. and Correa,B.M. : Human macrophage culture, the leprosy prognostic test (LPT). Int. J. Lepr., 35:277, 1967.
6. Bodla,D.M.S., Harry,E.B., Nagyan,S. and Kirchheimer, W.P. : Delayed hypersensitivity test with *Mycobacterium leprae* purified protein derivative. Lepr. India, 48:8, 1976.
7. Bolgiano,R. : Leprosy and genetics, a review of past research with remarks concerning future investigation. Bull. W H O, 57:461, 1967.
8. Bolgiano,R. and Picard,R.C.B. : Effect of D.D.S on phytohaemagglutinin induced lymphocyte transformation. Int. J. Lepr. 42:432, 1974.
9. Birdi,T.J., Salgme,P.R., Mahadevan,P.R. and Anita, N.H. : Role of macrophage in defective cell mediated immunity in lepromatous leprosy. II Macrophage and lymphocyte interaction. Int. J. Lepr., 48:175, 1980.

10.  Beck,A.A. and Macmillan,N.P. : The influence of leprosy on delayed type skin reactions and serum agglutination titres to common tubercle bacilli: comparative study of patients with lepromatous and tuberculoid leprosy and controls in Ethiopia. Am. J. Hyg., 77:505, 1963.
11.  Bullock,W.E. : Studies of immune mechanism in leprosy, depression of delayed allergic response to skin test antigens. N. Engl. J. Med., 270:290, 1964.
12.  Bullock,W.E., Calladine,M.L. and Tanner, B.J. : Immunohistologic alteration of skin and ultra-structural changes of glomerular basement membranes in leprosy. Am. J. Trop. Med. Hyg., 23:81, 1974.
13.  Bullock,W.E. and Faval,P. : Studies of immune mechanism in leprosy, the role of cellular and humoral factors in the impairment of the vitro immune response. J. Immunol., 106:699, 1971.
14.  Chogle,J.B., Khanolkar,S.R. and Anita,V.H. : T & B lymphocyte in the spectrum of leprosy. Lepr. India, 49:36, 1977.
15.  Glaman,H.M., Chaperon,S.A. and Tripplett,B.F. : Immunosepstone of transferred thymus marrow cell combination. J. Immunol., 97:689, 1966.
16.  Gurdakshik,R., Duguid,J.P., Morrison,R.P. and Austin, R.H.A. : Medical Microbiology Ed. 12th, Pub. Churchill Livingston, Great Britain, London, pp 804, 1975.
17.  Gulling,G.P.A. : Hand book of histopathological and histochemical techniques. Ed. 2nd., Pub. Butterworths, London, pp 397, 1974.
18.  Danis,J.V. and Lewis,S.W. : Practical haematology, Ed. V, Pub. Churchill Livingston, Edinburgh, pp 47, 1975.

29. David,J.R. and David,R.B. : Cellular hypersensitivity and immunity. Proc. Albany, 26:200, 1972.
30. Davis,A.J.S., Lounsbury,L., Wallis,V., Merchant,R. and Elliott,S.V. : The failure of thymus derived cells to produce antibody. Transplantation, 3:222, 1967.
31. Dhamondam : Studies of leprosin test. The active principles of leprosin is a protein antigen of the bacillus. Lepr. India, 43:69, 1961.
32. Dhamondam and Chatterjee,S.N. : A proposed system of classification of leprosy. Lepr. India, 43:242, 1963.
33. Dierck,R.E. and Shepard,G.C. : Effect of phytohaemagglutinin and various mycobacterial antigens on lymphocyte culture from leprosy patients. Proc. Soc. Exp. Med., 127:391, 1968.
34. Drutz,D.J. and Gutman,B.A. : Renal manifestation of leprosy, glomerulonephritis a complication of erythema nodosum leprosum. Am. J. Trop. Med., 23:496, 1972.
35. Dubey,S.K., Jogikar,V.K., Hardas,U.D. and Chaudhary, B.S. : A study of cell mediated immunity in leprosy. Lepr. India, 43:197, 1961.
36. Dwyer,J.M., Bullock,T.E. and Field,J.P. : Dieter-  
burnes in blood, T : B lymphocyte ratio in lepromatous leprosy, clinical and immunological correlations. N. Engl. J. Med., 286:1686, 1972.
37. Finsen,V.J.H.W. : The early reaction induced by leprosin. Int. J. Lept., 6:1, 1940.
38. Friedman,B.M., Wybran,J. and Robbins,D. : T-  
Rosa forming cells, cellular immunity and cancer. N. Engl. J. Med., 291:475, 1974.
39. Ganti-Venkatesh,K.J., Lin,S.P., Jacobson,R.R. and Good,R.A. : B-lymphocytes in lepromatous leprosy. N. Engl. J. Med., 286:1683, 1972.

30. Gellher, R.E., Brown, D.J., Spotska, W.L. and Fischl, P. : Clinical correlation of C<sup>3q</sup> precipitating substances in sera of patients with leprosy. Am. J. Trop. Med. Hyg. 23:471, 1974.
31. Chakraborty, S.K., Ganguly, U. and Banerji, G.: Phytobean agglutinin (PBA) - induced transformation of peripheral blood lymphocytes in leprosy patients. Leprosy India, 22:225, 1980.
32. Godal, T., Nykjaer, B., Gamble, H.H. and Myrvang, B. : Characterization of cellular immune defect in leprosy patients. Leprosy reactive lymphocyte. Clin. Exp. Immunol., 9:621, 1971.
33. Godal, T. : Immunological aspect of leprosy. Proc. Allergy, 25:211, 1976.
34. Greaves, M.F. and Brown, G. : A human B-lymphocyte specific antigen. Nature, 246:216, 1973.
35. Quinto, R.S., Mahaley, C.W. and Doull, A.J. : Cutaneous response to leprosin and to other mycobacterial antigens. Int. J. Lep., 30:183, 1968.
36. Quinto, R.S. : Skin tests in leprosy. Annals. N.Y. Acad. Med., 100:149, 1962.
37. Han, S.H., Weimer, R.G. and Kao, S.T. : Prolonged survival of skin allografts in leprosy patients. Int. J. Lep., 39:1, 1971.
38. Han, S.H., Weimer, R.G. and Lin, Y.C. : Transformation of leprosy lymphocyte by leprosin, tuberculolin and phytobean agglutinin. Int. J. Lep., 39:709, 1971.
39. Hansen, G.A. : Nord. Mag. Lægevidensk 6:1, 1974 ; reprinted in part, in English translation in Int. J. Lep., 22:307, 1980.

40. Marton, M., Glens, G., Bjørnstad, B., Bratvall, G. and Aunezen, N.K. : Mycobacterium leprae specific antibodies detected by radioimmune assay. *Scand. J. Immunol.*, 7:111, 1978.
41. Harris, T.H., Gyurek, L., Mortara, E. and Abrech, W.B. : The role of lymphocyte in antibody formation. *J. Exp. Med.*, 81:73, 1945.
42. I.C.M.R. : Status report on leprosy; Immunology of leprosy. Pub. I.C.M.R., New Delhi, pp 22, 1981.
43. VII International Congress of Leprology (Tokyo) : Technical resolution in immunology in transmission of VII International Congress of Leprology, Tokyo, Tokyo. Kyobai, pp 468, 1989.
44. Job, G.K., Chakraborty, S.J.G., Taylor, P.M., Danis, P.M. and Jasudian, G. : Evaluation of cell mediated immunity in histopathologic spectrum using lymphocyte transformation test. *Int. J. Lepr.*, 44:236, 1976.
45. Dondal, M., Helm, G. and Vignati, R. : Surface marker on human T and B lymphocytes. *J. Exp. Med.* 206:1207, 1977.
46. Kuklancik, L., Pranga, N., Kouznetzkoglova, N., Karalis, D., and Trichopoulou, D. : Cellular immunity in patients with leprosy. Circulating T-lymphocytes and their response to PHA in leprosy. *Int. J. Lepr.*, 65:941, 1977.
47. Kirribhaver, V.P. and Sanchari, R.M. : Leprosy susceptibility testing of armadillo. I. Cellular response to intradermally inoculated heat killed leprosy bacilli. *Microbes*, 7:121, 1978.
48. Kirribhaver, V.P. and Stoerz, E.C. : Attempts to establish the armadillo (*Dasyprocta novemtaeniata* Linnaeus) as a model for study of leprosy. I. Report of leprosyoid leprosy in an experimentally infected armadillo. *Int. J. Lepr.*, 59:693, 1971.

49. Knottall,G., Stanton,J.L. and Vojak,G.P. : Studies of mycobacterial antigen with especial reference to *Mycobacterium leprae*. *Infect. Immun.*, 15:1138, 1976.
50. Kumar,B., Kaur,S., Ganguly,N.K. and Shyam,S. : Cutaneous response to antigen and irritants in patients of leprosy. *Lepr. India*, 52:405, 1980.
51. Landsteiner,K., and Chase,M.V. : Experiments on transfer of cutaneous sensitivity to simple compound. *Proc. Soc. Exp. Biol. Med.*, 49:600, 1942.
52. Leifer,D.L. : Intradermal tests with mycobacterial substances and normal tissue suspension. *Int. J. Lepr.*, 36:53, 1968.
53. London,P.C. : The name "leprosy". *Am. J. Trop. Med. Hyg.*, 1:999, 1902.
54. Lie,H.P. : On leprosy in the Bible. *Lepr. Rev.*, 9:185 and 88, 1938.
55. Lin,S.D., Jacobson,R.R., Park,B.H. and Good,R.A. : Quantitative analysis of thymus derived lymphocyte response to phytohaemagglutinin in leprosy. *Int. J. Lepr.*, 43:95, 1975.
56. Lin,S.D., Kisselkell,D.P., Jacobson,R.R., Choi,Y.S. and Good,R.A. : Thymus dependent lymphocytes in peripheral blood in leprosy patients. *Infect. Immun.*, 9:894, 1970.
57. Lowe,J. : Comment on the history of leprosy. *Ind. Med. Gazette*, 77:600, 1968.
58. Mackaness,G.B. : The influence of immunologically committed lymphoid cells on macrophage activity in vivo. *J. Exp. Med.*, 129:973, 1969.
59. Mehra,V., Mehta,L.M., Rothman,R., Reitberg,I., Schlesinger,S.P. and Bloom,B.R. : Delineation of human T-cell sub set responsible for leproxin induced suppression in leprosy patient. *J. Immunol.*, 135:1133.

60. ✓ Mehra, V.L., Valver, S.P., Mallikarjun, K. and Shantanu, L.K. : Influence of chemotherapy and serum factors on the mitogenic response of peripheral leucocytes of leprosy patients to phytohaemagglutinin. *Clin. Exp. Immunol.*, 18:205, 1973.
61. ✓ Mendes, N.F., Kapurantych, S. and Mehta, N.G.S. : T and B lymphocyte in patient with lepromatous leprosy. *Clin. Exp. Immunol.*, 16:23, 1974.
62. ✓ Mendes, N.F., Tolani, N.S.A., Sylveira, M.P.A., Gilbertson, R.B. and Metagar, R.S. : Technical aspect of the rosette tests used to detect human complement receptor 'B' and sheep erythrocyte binding T-lymphocyte. *J. Immunol.*, 111:166, 1973.
63. ✓ Mehta, G., Chacko, C.J.G., Sunder Rao, P.S.S. and Job, C.K. : T-cell depletion in patients with long standing lepromatous leprosy. *Lepr. India*, 52:366, 1980.
64. ✓ Miller, J.P.A.P. and Mitchell, G.P. : Cell to cell interaction in immune response. *J. Exp. Med.*, 128:803, 1968.
65. ✓ Mitouda, K. : On the value of skin reaction to a suspension of skin nodule. *Jap. J. Dermatol. Ven.*, 19:697, 1919 ; English translation in *Int. J. Lep.*, 31:247, 1953.
66. ✓ Moyan, C.J., Ryder, G., Turk, J.L. and Waters, M.P.R. : Evidence for circulating immune complexes in lepromatous leprosy. *Lancet*, 10:572, 1972.
67. ✓ Nyveng, B., Gedal, P., Ridley, D.S., Proland, S.S. and Song, Y.H. : Immune responsiveness to *Mycobacterium leprae* and other mycobacterial antigen throughout the clinical and histopathological spectrum of leprosy. *Clin. Exp. Immunol.*, 14:581, 1973.

68. ✓ Nath, I., Curtis, J., Sharma, A.K. and Talwar, G.P. : Circulating T-cell numbers and their mitogenic potential in leprosy, correlation with mycobacterial load. *Clin. Exp. Immunol.*, 29:298, 1977.
69. ✓ Navelkar, R.G. : Immunologic analysis of *Myc. leprae* antigen by means of diffusion in gel method. *Int. J. Lepr.*, 39:105, 1971.
70. Nelson, D.S., Nelson, M., Thurston, J.W., Waters, M.P.R. and Pearson, J.M.H. : Phytohaemagglutinin induced lymphocyte transformation in leprosy. *Clin. Exp. Immunol.*, 9:23, 1971.
71. ✓ Ogilvie, H. Thomson, William, A.R. and Garland, J. : Biblical leprosy. *The Practitioner*, 177:665, 1956.
72. ✓ Piscani, R.C.B., Beignelman, D. and Opyonellin, D.V.A. : In vitro behaviour of blood derived macrophages against killed *M. leprae*. *Int. J. Lepr.*, 61:14, 1973.
73. ✓ Puri, Ching Wong, Chan Yeeh, C.H., Wu, S. and Kendall, P.H. : Transformation of lymphocyte by PHA in leprosy sera. *Int. J. Impr.*, 39:7, 1971.
74. ✓ Quiseric, P.P., Rao, T.H., Ioven, N.S. and Prion, G.J. : Immunoglobulin deposits in lepromatous leprosy skin. *Dermatol.* 111:331, 1975.
75. ✓ Raballo, P.S. and Aszkenasy, R.D. : Immunological principles as a guide to new leprosy concept. A life long study. *Int. J. Dermatol.* 14:770, 1975.
76. ✓ Rao, S.S.L. and Rao, P.R. : Immunological status of nonleprosy leprosy. Leucocyte migration inhibition test as a measure of cell mediated immune response. *Lepr. India*, 53:240, 1971.
77. ✓ Rao, T.H., Quiseric, P.P., Harding, D., Heis, K.N., Diazia, P.J., Lenn, L.F. and Prion, G.J. : Intradermal antigen, epicutaneous haptens, T-cell count, B-cell count, lymphocyte transformation test and auto anti-

- bedies in one group of patient with lepromatous leprosy. Int. J. Lepr. 42:269, 1974.
78. Rao,T.M. and Iyer,N.S. : Erythema nodosum leprosy in a general hospital. Arch. Dermatol., 111:1572, 1975.
79. Rao,T.M., Quimioio,P.P. and Harding,B. : Immunologic responses in patients with lepromatous leprosy. Arch. Dermatol., 112:791, 1976.
80. Ridley,D.S. : Review of five group system for the classification of leprosy according to immunity. Int. J. Lepr., 40:103, 1972.
81. Ridley,D.S. and Jopling,W.H. : A classification of leprosy for research purposes. Lepr. Rev., 33:119, 1962.
82. Ridley,D.S. and Jopling,W.H. : Classification of leprosy according to immunity - A five group system. Int. J. Lepr., 34:255, 1966.
83. Ridley,D.S. and Waters,M.F.R. : Significance of variation within the lepromatous group. Lepr. Rev., 40:643, 1969.
84. Rogers,L. and Muir,E. : Leprosy second Ed. John Wright, Bristol, pp 1, 1940.
85. Reitt,I.N., Greaves,M.F., Torrigiani,G., Brostoff,J. and Playfair,J.H.L. : The cellular basis of immunological responses. Lancet, ii:1267, 1969.
86. Rojas-Sopena,G., Mondon-Mavarroto,I. and Estrada-Perez,S. : Phenome of C<sup>3</sup>q - reactive immune complexes in patients with leprosy. Clin. Exp. Immunol., 13:218, 1972.
87. Roslands,P.T. and Dennis,R.P. : Surface receptor in immune responses. N. Eng. J. Med., 293:126, 1975.

88. Sandhu,K.N., Mathur,B.B. and Chawla,S.N. : Status of circulating T-lymphocyte population in leprosy. Lepr. India, 81:199, 1980.
89. Saha,E. and Mittal,M.M. : A study of cell mediated immunity in leprosy, changing trends in immunological spectrum of the disease. Clin. Exp. Immunol., 9:901, 1971.
90. Scott,H.H. : The influence of the slave trade in the spread of tropical disease. Trans. Roy. Soc. Med. and Hyg., 37:169, 1943, (Quoted in 'Leprosy' Vol. I by Dharmsena Ed. I, Pub. Kothari Med. Pub. House, Bombay pp 7, 1978).
91. Seligman,M.B. : B-cell and T-cell marker in lymphoid proliferation. N. Engl. J. Med. 290:1489, 1974.
92. Sen Gupta,U., Ph.D., Senior Research Officer : On personal communication, central JAIIB Institute for Leprosy, Agra, 1981.
93. Sharma,S., Ganguly,N.K., Kumar,B., Kaur,S. and Chakraverty,R.N. : T and B lymphocyte and blastogenesis in leprosy. Lepr. India, 81:194, 1979.
94. Shree,T. : Immune complexes in glomeruli of patients with leprosy. Lepr. Rev., 43:282, 1972.
95. Sinha,S. and Sen Gupta,U. : Assessment of Dharmsena antigen (IV) antigenic analysis of leprosy. Lepr. India, 83:6, 1981.
96. Stanford,J.L. and Rock,G.A.W. : Taxonomic studies on the leprosy bacillus. Int. J. Lepr., 44:216, 1976.
97. Tekmar,G.P., Krishna,A.D., Nehru,V.L., Blum,L.A. and Peterson,J.M.H. : Evaluation of cell mediated immune response in untreated cases of leprosy. Clin. Exp. Immunol., 18:195, 1972.

98. Turk,J.L. and Waters,M.P.R. : Cell mediated immunity in patients with lepromatous leprosy. *Lancet*, II: 363, 1969.
99. Turk,J.L. and Waters,M.P.R. : Immunological significance of changes in lymphnodes across the leprosy spectrum. *Clin. Exp. Immunol.* 6:363, 1971.
100. Virch,W., Salas,B. de. and Condit,J. : Lymphocyte transformation with phytohaemagglutinin in leprosy. *Int. J. Lepr.*, 40:4, 1972.
101. Verma,R.C., Balkrishnan,K., Van der Veen,D.M. and Talwar,G.P. : Lymphocytic bearing immunoglobulin determinant in normal human lymphode and patients with lepromatous leprosy. *Int. J. Lepr.*, 29:90, 1971.
102. Wade,H.H. : The classification of leprosy - A proposed synthesis based primarily on the Rio de Janeiro - Havana system. *Int. J. Lepr.*, 20:429, 1982.
103. Yamada,S.N.G., Turk,J.L., Waters,M.P.R. and Ross, R.J.W. : Erythema nodosum leprosum - a clinical manifestation of arthus phenomenon. *Lancet*, II : 933, 1969.

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**"STUDY OF CELL MEDIATED IMMUNITY IN IMPROVE"**

**Investigator : PREM KUMAR SINGH**

**CASE PROFORMA**

**Case No.:**

**H.R.D. No.:**

**O.P.D. No.:**

**Patient's Name:** **Age/Sex:** **Ward/Bed:**

**Clinical Diagnosis:** **Physician:**

**Socio-economic status:**

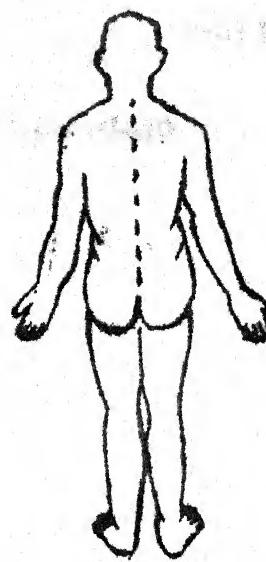
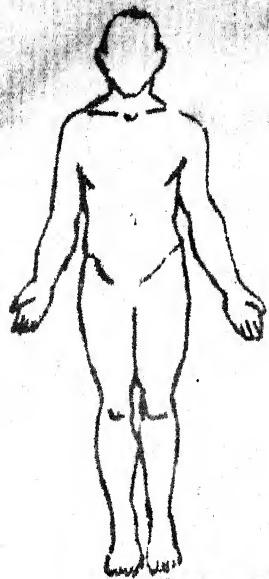
**Complaints of :** **Duration:**

**Family History:**

**General Examination :**

- Built
- Eye Disease
- Bridge of Nose
- Lymphnodes

- Negroes
- Hand & Foot
- Miscellaneous

**Systemic Examination:**a) General:**Lab. Findings :****1. Blood :-      Blood group.**

T.L.G.

D.L.G.

Hb%

G.S.B.

G.B.P.

**2. a) Hematology (A.P.S.) :****b) Skin Survey (A.P.S.) :****3. Misstoxicological findings :**

• Skin

• Other tissues

• A.P.S.

4. a) Absolute Lymphocyte Count

b) T-cell Count -

T-cell %

c) B-cell Count -

B-cell %

5. Blast transformation percentage :

6. a) lepromatous Test :      48 Hours      21 Days

b) Candida Test :

Abbreviations used

AFB	:	Acid Fast Bacilli
ALC	:	Absolute Lymphocyte count
BB	:	Borderline
B-cell	:	Bone equivalent derived cell
BL	:	Borderline lepromatous
BT	:	Borderline tuberculoid type
C	:	Control
CMI	:	Cell Mediated Immunity
Con.A	:	Concanavalin A
DNCB	:	Di Nitro Chloro Benzene
EAC rosette:	:	Erythrocyte antibody complement rosette
EML	:	Erythema Nodosum Leprosus
E-rosette:	:	Erythrocyte rosette
ICMR	:	Indian Council of Medical Research
LI	:	Leprosy Indefinite
LMT	:	Lymphocyte Migration Inhibition Test
LTT	:	Lymphocyte Transformation Test
MEN	:	Minimum Essential Medium
Mycob. lepros:	:	Mycobacterium leprae
MLT	:	Human Lymphocyte Transfer Test
PBS	:	Phosphate Buffer Saline
PTA	:	Phytohaemagglutinin
SD	:	Standard Deviation
SBC	:	Sheep Red Blood Cell
T-cell	:	Thymus derived cell
T C Medium 199	:	Tissue Culture Medium 199
TI	:	Tuberculoid Indefinite
TLC	:	Total Lymphocyte Count
TT	:	Tubercle Type